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## PROVISIONAL APPLICATION FOR PATENT COVER SHEET

This is a request for filing a PROVISIONAL APPLICATION FOR PATENT under 37 CFR 1.53(c).

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This collection of information is required by 37 CFR 1.51. The information is used by the public to file (and by the PTO to process) a provisional application. Confidentiality is governed by 35 U.S.C. 122 and 37 CFR 1.14. This collection is estimated to take 8 hours to complete, including gathering, preparing, and submitting the complete provisional application to the PTO. Time will vary depending upon the individual case. Any comments on the amount of time you require to complete this form and/or suggestions for reducing this burden, should be sent to the Chlef Information Officer, U.S. Patent and Trademark Office, U.S. Department of Commerce, Washington, D.C. 20231. DO NOT SEND FEES OR COMPLETED FORMS TO THIS ADDRESS. SEND TO: Box Provisional Application, Assistant Commissioner for Patents, Washington, D.C. 20231.

### PROVISIONAL APPLICATION COVER SHEET Additional Page

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Number 1 of 1

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File No.: 3110 0017

April 15, 2002

### **DELIVERED BY HAND**

Assistant Commissioner for Patents
USPTO Office of Initial Patent Examination
Crystal Plaza Building 2, Room
2011 South Clark Place
Arlington, Virginia 22202
U.S.A.

Dear Sir.

Re:

**New United States Provisional Patent Application** 

Title:

**TB VACCINES** 

Inventors:

LIU, J., CHEN, J., and ALEXANDER, D.C.

We apply in the name of J. Liu, J. Chen, and D.C. Alexander for a provisional patent application entitled TB VACCINES.

In addition to the \$150.00 filing fee, included in our firm cheque, we enclose the following documents:

- 1. provisional application cover page; and
- 2. patent application

Please direct any questions to Kathryn Schubert at 416-941-9027.

Yours very truly,

Gervas W\Wall

Registration No. 35766

GWW/KS/el Encl. Tuberculosis Vaccines Including Recombinant BCG Strains Expressing Alanine Dehydrogenase, Serine Dehydratase and/or Glutamine Synthetase

### Field of the Invention

This invention relates to tuberculosis (TB) vaccines.

### **Background of the Invention**

TB is a deadly contagious disease caused by the infectious agent, Mycobacterium tuberculosis. It kills 2 million people each year. The World Health Organization (WHO) 2001 annual report estimated that there would be 8.4 million new TB cases in 1999, up from 8.0 million in 1997. If the present trend continues, it is estimated that between 2000 and 2020, nearly one billion people will be newly infected, 200 million people will become ill and 35 million will die from TB. The spread of HIV/AIDS and the emergence of multidrug-resistant TB contribute to the worsening impact of this disease. Bacille Calmette-Guérin (BCG), an attenuated strain of Mycobacterium bovis, is currently the only available vaccine for the prevention of TB. In animal models of infection, BCG vaccination has been demonstrated to induce protective immunity against a M. tuberculosis challenge (Baldwins et al., 1998). In humans, BCG vaccination has demonstrated consistent protection against the childhood forms of TB, especially meningitis. However, BCG vaccination is controversial due to variations in its efficacy for protecting adults from pulmonary TB (Fine, 1989; Colditz et al., 1994; Sterne et al., 1998). Trials conducted in the 1940s and 1950s in developed countries such as the United Kingdom, Denmark and North America demonstrated the vaccine to be highly efficient (70-80%). However, in the single largest clinical trial, which took place in India in 1970s and involved more than 265,000 persons, BCG vaccination provided no detectable protection against pulmonary TB. Thus, there is an urgent need to generate an improved vaccine(s) to replace the BCG and to prevent TB.

Several explanations have been suggested for the variation in protective efficacy of BCG (Andersen, 2001). The most prominent hypothesis is that exposure to environmental mycobacteria sensitizes the host against mycobacteria in general, thereby providing

heterologus immunity that obscures the potential benefits of BCG vaccination (Fine, 1995; Fine and Vynnycky, 1998). Furthermore, a recent study showed that the multiplication of BCG was inhibited in animals sensitized with environmental mycobacteria, and consequently BCG vaccination elicited only a transient immune response and failed to provide protective immunity against TB (Brandt et al., 2002). This study also supports the long-standing observation that the induction of immunity to TB requires productive infection by BCG. BCG is a live vaccine; killed BCG does not provide protection. Like M. tuberculosis, BCG is capable of forming granulomas and abscesses in various tissues in the infected host (Hogan et al., 2001). The ability of M. tuberculosis and M. bovis BCG to survive and persist within granulomas, a hostile environment with restricted access to nutrients and reduced oxygen tension, appears to be dependent on the ability of the bacteria to adapt their metabolism to the available source of carbohydrate, nitrogen, and energy (Barclay and Wheeler, 1989). A recent study revealed that fatty acids serve as a source of carbohydrates and are required for persistence of M. tuberculosis in mice and activated macrophages (McKinney et al., 2000). Following vaccination in immunocompetent individuals, BCG may persist for certain periods before it is eliminated from the host (Dunn and North 1995; Lagranderie et al., 1996; Moisan et al., 2001).

The key to developing a new and effective TB vaccine is to provide long-term protection (Orme, 2001; Young, 2000). Existing BCG vaccines impart protection against the manifestations of TB in children, but their efficacy wanes over a period of 10 to 15 years, presumably because the protective immunity induced by BCG is gradually lost (Orme, 2001). New strategies to developing an improved vaccine have included the use of attenuated mycobacteria, subunit vaccines and DNA vaccines (Andersen, 2001). However, none of these have proved to be more potent than, or even as effective as BCG. Survival and growth of *M. bovis* BCG is necessary for eliciting protective immunity. It has been shown that early treatment of infected mice with isoniazid to inhibit bacillary growth prevents the development of acquired resistance. BCG strains that persist for extended periods within the host are required in order to obtain more effective vaccines. As such, there is a need for novel, recombinant strains of Bacille Calmette-Guérin.

### **Summary of the Invention**

The invention provides vaccines that overcome the limited ability of BCG strains to use naturally occurring amino acids as the nitrogen source for growth. Furthermore, L-alanine, D-alanine, or L-serine inhibits the growth of BCG strains even when ammonium is present. Expressing a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2] in BCG strains relieves the growth inhibition of BCG by alanine. Similarly, expressing a functional L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6] in BCG strains relieves the growth inhibition of BCG by L-serine. The mechanism for such inhibition occurs through blockage of glutamine synthetase. Overexpression of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO:14] in BCG relieves the growth inhibition of BCG by alanine and L-serine. Recombinant BCG strains that express (or overexpress) a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6], and/or glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO:14] survive and persist longer within the host and consequently induce long-term protective immunity. Such persistent recombinant BCG strains provide more effective vaccines for the prevention of TB and other mycobacterial infections.

The present invention relates to recombinant *Mycobacterium bovis* BCG, which express DNA encoding an alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO: 2], a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6], and/or a glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO:14]. We found that, due to the lack of a functional alanine dehydrogenase [SEQ ID NO:3; SEQ ID NO:4], BCG cannot utilize alanine (L-alanine or D-alanine) as the only nitrogen source for growth. We further found that alanine (L-alanine or D-alanine) inhibits the growth of all BCG vaccine strains. Said inhibition is relieved by expressing a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2] in BCG. Similarly, BCG cannot utilize L-serine as the only nitrogen source for growth and that growth of BCG is inhibited by L-serine. Expressing a L-serine dehydratase [SEQ ID NO:6] in BCG strains relieves the growth inhibition by L-serine.

Alanine (L-alanine or D-alanine) and L-serine inhibits BCG growth likely by blocking the activity of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14]. Overexpression of

glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] in BCG relieves the growth inhibition of BCG by alanine and L-serine. Glutamine synthetase, in conjunction with glutamate synthase, provides glutamine and glutamate, which are essential for biosynthesis of all amino acids, proteins, purines and pyrmidines. Inhibition of glutamine synthetase stops cell growth. Supplying amino acids that can be converted to glutamate such as L-glutamine, L-glutamate, L-aspartate, and L-asparagine can relieve such inhibition. Indeed, our data show that the inhibition of BCG growth by alanine (L-alanine or D-alanine) or L-serine is relieved by supplementing growth medium with L-glutamine, L-glutamate, L-aspartate, or L-asparagine.

Since BCG is a live vaccine, recombinant BCG strains expressing or overexpressing a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO: 2], a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO: 6], and/or a glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] survive longer within the human host and subsequently induce long-term memory immunity. These recombinant BCG strains provide extremely useful vaccines.

The present invention relates to a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide that exhibits alanine dehydrogenase activity [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase activity [SEQ ID NO:7 to SEQ ID NO:14], or L-serine dehydratase activity [SEQ ID NO:5; SEQ ID NO:6].

The invention also relates to a live recombiant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide selected from the group consisting of alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase [SEQ ID NO:7 to SEQ ID NO:14] and L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6].

The invention further relates to a live recombinant Mycobacterium bovis-BCG strain comprising a nucleic acid capable of expression, the nucleic acid comprises all or part of at least one nucleic acid molecule selected from the group consisting of [SEQ ID NO:1], [SEQ ID NO:5], [SEQ ID NO:6], [SEQ ID NO:7], [SEQ ID NO:8],

[SEQ ID NO:9], [SEQ ID NO:10], [SEQ ID NO:11], [SEQ ID NO:12], [SEQ ID NO:13] and [SEQ ID NO:14].

In one embodiment, the live recombinant Mycobacterium bovis-BCG strain is selected from the group consisting of Mycobacterium bovis-BCG-Russia, Mycobacterium bovis-BCG-Moreau, Mycobacterium bovis-BCG-Japan, Mycobacterium bovis-BCG-Sweden, Mycobacterium bovis-BCG-Birkhaug, Mycobacterium bovis-BCG-Prague, Mycobacterium bovis-BCG-Glaxo, Mycobacterium bovis-BCG-Denmark, Mycobacterium bovis-BCG-Tice, Mycobacterium bovis-BCG-Frappier, Mycobacterium bovis-BCG-Connaught, Mycobacterium bovis-BCG-Phipps, and Mycobacterium bovis-BCG-Pasteur.

Another aspect of the invention is a pharmaceutical composition comprising a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide that exhibits alanine dehydrogenase activity [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase activity [SEQ ID NO:7 to SEQ ID NO:14], or L-serine dehydratase activity [SEQ ID NO:5; SEQ ID NO:6].

The invention also relates to a live recombiant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide selected from the group consisting of alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase [SEQ ID NO:7 to SEQ ID NO:14] and L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6].

In yet another aspect of the invention there is a pharmaceutical composition comprising a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid comprises all or part of at least one nucleic acid molecule selected from the group consisting of [SEQ ID NO:1], [SEQ ID NO:2], [SEQ ID NO:5], [SEQ ID NO:6], [SEQ ID NO:7], [SEQ ID NO:8], [SEQ ID NO:9], [SEQ ID NO:10], [SEQ ID NO:11], [SEQ ID NO:12], [SEQ ID NO:13] and [SEQ ID NO:14].

In a futher aspect of the invention there is a vaccine or immunogenic composition for treatment or prophylaxis of a mammal against challenge by mycobacteria comprising a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide that exhibits alanine dehydrogenase activity [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase activity [SEQ ID NO:7 to SEQ ID NO:14], or L-serine dehydratase activity [SEQ ID NO:6].

In another aspect of the invention there is a vaccine or immunogenic composition for treatment or prophylaxis of a mammal against challenge by mycobacteria comprising a live recombiant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide selected from the group consisting of alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase [SEQ ID NO:7 to SEQ ID NO:14] and L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6].

In yet another aspect of the invention there is a vaccine or immunogenic composition for treatment or prophylaxis of a mammal against challenge by mycobacteria comprising a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid comprises all or part of at least one nucleic acid molecule selected from the group consisting of [SEQ ID NO:1], [SEQ ID NO:2], [SEQ ID NO:5], [SEQ ID NO:6], [SEQ ID NO:7], [SEQ ID NO:8], [SEQ ID NO:9], [SEQ ID NO:10], [SEQ ID NO:11], [SEQ ID NO:12], [SEQ ID NO:13] and [SEQ ID NO:14]. In a preferred embodiment the vaccine or immunogenic composition is for the treatment or prophylaxis of a mammal against challenge by *Mycobacterium tuberculosis*. In another preferred embodiment the vaccine or immunogenic compositions of the current invention further comprise a pharmaceutically acceptable carrier. In yet another preferred embodiment the vaccine or immunogenic compositions further comprise adjuvants. In a another embodiment the vaccine or immunogenic compositions further comprises immunogenic materials from one or more other pathogens.

Another aspect of this invention relates to a method for treatment or prophylaxis of a mammal against challenge by *Mycobacterium tuberuclosis* or *Mycobacterium bovis* comprising administering to the mammal a vaccine or immunogenic composition of the instant invention. In one embodiment the mammal is a cow. In another embodiment the mammal is a human. In yet another embodiment the vaccine or immunogenic composition is administered in the presence of an adjuvant.

A further aspect of the invention is a method for the treatment or prophylaxis of a mammal aginst cancer comprising administering to the mammal a vaccine or immunogenic composition of the current invention. In one embodiment the cancer is bladder cancer. In another embodiment the vaccine or immunogenic composition is administered in the presence of an adjuvant.

The invention also relates to a test kit comprising the live recombinant *Mycobacterium* bovis-BCG strain of the instant invention.

The invention further relates to a media composition for inhibiting the growth of *Mycobacaterium bovis*-BCG comprising alanine as the only nitrogen source for growth. In another embodiment serine is the only nitrogen source for growth. In another embodiment, the media compositions of the current invention further comprise a carbon source, iron, magnesium, and SO<sub>4</sub>. In one embodiment the carbon source is selected from the group consisting of glycerol, dextrose, citrate, and glucose.

The current invention relates to a method for inhibiting the growth of *Mycobacterium* bovis-BCG comprising the steps of (a) obtaining a sample comprising *Mycobacterium* and (b) culturing the sample in a selective media. In one embodiment the selective media comprises alanine as the only nitrogen source. In yet another embodiment the selective media comprises serine as the only nitrogen source.

Another aspect of the invention relates to a method for culturing Mycobacterium bovis-BCG comprising the steps of (a) obtaining a sample comprising Mycobacterium and (b)

culturing the sample in differential media. In one embodiment the differential media comprises histidine.

### **Brief Description of the Drawings**

Preferred embodiments of the invention will be described in relation to the drawings in which:

Fig. 1. Cloning of the ald gene. First, a 4.5 kb ScaI fragment of M. tuberculosis genomic DNA containing the ald gene [SEQ ID NO:1] was ligated to Ecl136II-linearized pUC19 to generate pUC-ALD. Then, mycobacterial plasmid pALD was created by ligating the 1.9 kb KpnI fragment containing the ald gene [SEQ ID NO:1] to KpnI-linearized pMD31.

### Fig. 2. Cloning of the sdaA gene.

Cloning of sdaA [SEQ ID NO:5] was accomplished in two steps. First, a 9.5 kb BamHI fragment of M. tuberculosis genomic DNA was ligated to BamHI-linearized pMD31 to generate pSDA1. Plasmid pSDAA was generated by cleavage of pSDA1 with PstI, followed by self-ligation of the 10.9 kb PstI fragment.

Fig. 3. Inhibition of BCG growth by L-alanine in GAS. BCG-Japan, BCG-Frappier, and BCG-Pasteur grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media, were each inoculated into duplicated 5 ml culture volumes of GAS, GAS without L-alanine, and GAS supplemented with 27 mM L-asparagine, to a cell density of 2 ×10<sup>7</sup> cells/ml. Cultures were incubated at 37°C with constant shaking for 16 days and then 2 ml aliquots of cell culture were centrifuged and cell pellet lyophilized to determine cell dry weight.

Fig. 4. Inhibition of BCG growth by increasing concentrations of L-alanine in Sauton containing NH<sub>4</sub>Cl (5 g/liter). a) BCG-Japan, b) BCG-Frappier, and c) BCG-Pasteur, grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media. Cells were washed and resuspended in Sauton basal medium (no nitrogen source).

Resuspended cells of each strain were inoculated into duplicate 5 ml culture volumes of Sauton media supplemented with NH<sub>4</sub>Cl and increasing concentrations of L-alanine. Cultures were incubated at 37°C with constant shaking for 30 days and cell dry weight was determined.

Fig. 5. Inhibition of BCG growth by D-alanine in GAS. BCG-Japan, BCG-Frappier, and BCG-Pasteur grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media, were each inoculated into 5ml culture volumes of GAS in which L-alanine was replaced by D-alanine, GAS without L-alanine and, GAS (containing D-alanine) supplemented with 27 mM L-asparagine, to a cell density of 2 ×10<sup>7</sup> cells/ml. Cultures were incubated at 37°C with constant shaking for 13 days and cell dry weight was determined.

Fig. 6. Growth of recombinant BCG strains expressing alanine dehydrogenase [SEQ ID NO:1] in GAS medium. The growth of BCG-Frappier/ald, BCG-Pasteur/ald, BCG-Frappier/pMD31, BCG-Pasteur/pMD31, BCG-Frappier, and BCG-Pasteur were compared. Cells of each strain, grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media, were washed and resuspended in Sauton basal medium (no nitrogen source). Resuspended cells were inoculated into duplicate 5 ml culture volumes of GAS without L-alanine, GAS containing L-alanine and GAS in which L-alanine was replaced by D-alanine. Cultures were incubated at 37°C with constant shaking for 15 days and cell dry weight was then determined.

Fig. 7. Inhibition of BCG growth by L-serine in GAS. BCG-Japan, BCG-Frappier, and BCG-Pasteur grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media, were each inoculated into duplicate 5 ml culture volumes of GAS in which L-alanine was replaced by L-serine, GAS without L-alanine, and GAS (containing L-serine) supplemented with 27 mM L-asparagine, to a cell density of 2 ×10<sup>7</sup> cells/ml. Cultures were incubated at 37°C with constant shaking for 15 days and cell dry weight was then determined.

Fig. 8. Growth of recombinant BCG strains expressing L-serine dehydratase [SEQ ID NO:5] in GAS medium containing L-serine. The growth of BCG-Japan/sdaA,

BCG-Frappier/sdaA, BCG-Pasteur/sdaA, BCG-Japan, BCG-Frappier, and BCG-Pasteur were compared. Cells of each strain, grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media, were washed and resuspended in Sauton basal medium (no nitrogen source). Resuspended cells were inoculated into duplicate 5 ml culture volumes of GAS without L-alanine, GAS in which L-alanine was replaced by L-serine, and GAS (containing L-serine) supplemented with 27 mM L-asparagine. Cultures were incubated at 37°C with constant shaking for 15 days and cell dry weight was then determined.

Fig. 9. Alignment of A) nucleotide and B) amino acid sequences of the ald genes of Mycobacterium tuberculosis (M. tb) [SEQ ID NO:1; SEQ ID NO:2] and Mycobacterium bovis (M. bovis) [SEQ ID NO:3; SEQ ID NO:4]. The point deletion causing the frameshift mutation in M. bovis ald [SEQ ID NO:3] is indicated with an arrow. Nucleotide codons and amino acids affected by this mutation are highlighted.

### **Detailed Description of the Invention**

BCG vaccine strains have a limited ability to utilize amino acids as the nitrogen source for growth. Furthermore, we found that naturally occurring amino acids L-alanine and L-serine inhibit the growth of BCG strains. Expressing a functional L-alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2] in BCG relieves the growth inhibition by alanine. Expressing of a functional L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6] in BCG relieves the growth inhibition by L-serine. As well, overproduction of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] relieves the growth inhibition by alanine and serine. These novel findings are significant because recombinant BCG strains that express (or overexpress) a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6], and/or glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] will survive better within the human host, induce long-term memory immunity and provide for more effective vaccines to prevent TB, particularly for protecting against pulmonary TB in adults.

It has long been known that administration of killed BCG strains results in a weak and transient immune response. Protective immunity requires survival and replication of BCG

in the vaccinated host. This notion is reinforced by a recent study of an animal model of infection, which showed that prior exposure to live environmental mycobacteria blocked the multiplication of BCG in infected mice. Consequently BCG elicited only a transient immune response which failed to provide protective immunity against TB (Brandt et al., 2002). Live BCG continuously secrete many different antigens that are likely important for the induction of protective immunity. The continuous production of numerous antigens by multiplying BCG gives live vaccines an advantage over subunit vaccines or DNA vaccines which transiently produce a few antigens. Thus the ability of BCG to multiply and persist within the host is an important determinant of BCG efficacy.

In order to grow and persist within the host, BCG must be able to utilize the available nutrients inside the host. It was demonstrated that isocitrate lyase, an essential enzyme for catabolism of fatty acids, is required for persistence of M. tuberculosis during the chronic phase of infection and that this requirement was dependent on an intact immune response of the host (McKinney et al., 2000). In another study, an M. bovis BCG strain lacking anaerobic nitrate reductase, an enzyme essential for nitrate respiration, failed to persist in lungs, liver and kidneys of immune-competent mice (Fritz et al., 2002). Our findings, that BCG strains utilize only a few types of amino acids as the nitrogen source for growth, and that the growth of all BCG strains are inhibited by naturally occurring L-alanine and L-serine, suggest that the ability of BCG to grow and persist within the host is restricted. The concentration of L-alanine that is available to BCG growing in human is estimated to be 0.33-0.42 mM (Barclay and Wheeler, 1989), which is sufficient to inhibit the growth of BCG-Pasteur or BCG-Frappier, and significantly reduce the growth of BCG-Japan (Fig. 4). The concentration of L-serine present in the extracellular fluids of the host is around 0.1 mM (Barclay and Wheeler, 1989), which may cause significant inhibition of BCG growth. Since multiplication of BCG is required to generate protective immunity, such inhibition by amino acids within the host may prevent the development of long-term protective immunity and hence the lack of protection against pulmonary TB in adults.

M. bovis BCG is also used in the treatment of bladder cancer. Numerous randomized controlled clinical trials indicate that intravesical administration of BCG can prevent or delay tumour recurrence (reviewed in Lamm, 2000; Lockyer and Gillatt, 2001). The

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details of how BCG exerts this effect remain to be determined. However, the antitumour response requires an intact T-cell response, and involves increased expression of Th1-type cytokines, including TNFa and IL-6 (reviewed in Prescott et al, 2000). The most effective treatment regimes involve multiple applications of BCG, which suggests that prolonged exposure to the bacteria is required. Similarly, tumours that retain the ability to phagocytize BCG are most susceptible to this treatment (de Boer et al 1996), indicating that bacterial interactions with the tumour are important. As such, a BCG strain demonstrating increased persistence may provide enhanced antitumour activity.

We show that the absence of a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2] is responsible for the failure of BCG strains to utilize alanine (L-alanine or Dalanine) as the only nitrogen source. A gene (Rv2708) coding for a L-alanine dehydrogenase (ald) [SEQ ID NO:1] was identified in the genome of M. tuberculosis. The activity of this enzyme from M. tuberculosis had been demonstrated biochemically in vitro. Ald converts L-alanine to pyruvate and ammonium, and is highly specific for Lalanine (Hutter and Singh, 1999). This enzyme was detected in the culture supernatent fraction of M. tuberculosis but not in M. bovis BCG-Japan nor BCG-Copenhagen, even though DNA Southern blot showed that the gene is present in both BCG strains (Anderson et al., 1992). Similarly, we do not detect alanine dehydrogenase activity in any of the 12 BCG strains listed in this report (data not shown). This lack of a functional alanine dehydrogenase in BCG strains is probably caused by a mutation within the ald gene [SEQ ID NO:3], and probably originated with the original M. bovis strain. A frame-shift mutation is found within the ald gene in the published genome sequence of M. bovis (Fig. 9) [SEQ ID NO:3]. As a result, the full length L-alanine dehydrogenase protein [SEQ ID NO:2; SEQ ID NO:4] cannot be made in BCG strains and subsequently BCG cannot catabolize alanine. Similarly, the failure of BCG to utilize L-serine as the only nitrogen source is likely to be caused by either mutations or altered expression of the sdaA gene [SEQ ID NO:5; SEQ ID NO:6], which encodes L-serine dehydratase. Expression of sdaA [SEQ ID NO:5; SEQ ID NO:6] of M. tuberculosis in BCG allows BCG strains to grow on L-serine as the only nitrogen source and relieves the inhibition of BCG growth by L-serine (Fig. 8). The inhibition of BCG growth by alanine and serine is

caused by inhibition of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14]. Overexpression of a glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] in BCG relieves the growth inhibition by L-serine, L-alanine and D-alanine.

BCG-Frappier and BCG-Pasteur are more susceptible than BCG-Japan to inhibition by alanine, presumably due to difference in the expression level or activity of glutamine synthetase. BCG-Japan differs from BCG-Frappier or BCG-Pasteur genetically (Behr et al., 1999). Calmette and Guérin developed the BCG vaccine in 1921 after 13 years and 230 passages of an isolate of *M. bovis in vitro*. Starting from 1924, BCG lots were distributed to laboratories around the world. These laboratories continued the passage of the bacteria *in vitro* employing a variety of different recipes and protocols until 1961when lyophilized seeds were established. As a consequence of such practices, different BCG progeny strains were created, which differed biochemically and genetically (Oettinger et al., 1999; Behr et al., 1999). Our data show that the ability of BCG strains to utilize amino acids as nitrogen source vary; for example, BCG-Japan is able to grow on cationic amino acids including L-arginine and L-lysine while BCG-Pasteur and BCG-Frappier cannot. These differences may also contribute to the differences of BCG efficacy in various clinical trials.

In summary, we use recombinant BCG strains that express (or overexpress) a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6], and/or glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] as vaccines to prevent TB and other mycobacterial infections. These recombinant BCG vaccines will induce long-term protective immunity against TB.

### Variations of Nucleic Acid Molecules

### **Modifications**

Many modifications may be made to the nucleic acid molecule DNA sequences disclosed in this application and these will be apparent to one skilled in the art. The invention includes nucleotide modifications of the sequences disclosed in this application (or fragments thereof) that are capable of directing expression in bacterial or mammalian

cells. Modifications include substitution, insertion or deletion of nucleotides or altering the relative positions or order of nucleotides.

### Sequence Identity

The nucleic acid molecules of the invention also include nucleic acid molecules (or a fragment thereof) having at least about: 70% identity, at least 80% identity, at least 90% identity, at least 95% identity, at least 96% identity, at least 97% identity, at least 98% identity or, most preferred, at least 99% or 99.5% identity to a nucleic acid molecule of the invention and which are capable of expression of nucleic acid molecules in bacterial or mammalian cells. Identity refers to the similarity of two nucleotide sequences that are aligned so that the highest order match is obtained. Identity is calculated according to methods known in the art. For example, if a nucleotide sequence (called "Sequence A") has 90% identity to a portion of [SEQ ID NO: 1], then Sequence A will be identical to the referenced portion of [SEQ ID NO: 1] except that Sequence A may include up to 10 point mutations (such as substitutions with other nucleotides) per each 100 nucleotides of the referenced portion of [SEQ ID NO: 1].

Sequence identity (each construct preferably without a coding nucleic acid molecule insert) is preferably set at least about: 70% identity, at least 80% identity, at least 90% identity, at least 95% identity, at least 96% identity, at least 97% identity, at least 98% identity or, most preferred, at least 99% or 99.5% identity to the sequences provided in SEQ ID NO:1 to SEQ ID NO:14 or its complementary sequence). Sequence identity will preferably be calculated with the GCG program from Bioinformatics (University of Wisconsin). Other programs are also available to calculate sequence identity, such as the Clustal W program (preferably using default parameters; Thompson, JD et al., Nucleic Acid Res. 22:4673-4680), BLAST P, BLAST X algorithms.

### Hybridization

The invention includes DNA that has a sequence with sufficient identity to a nucleic acid molecule described in this application to hybridize under stringent hybridization conditions (hybridization techniques are well known in the art). The present invention

also includes nucleic acid molecules that hybridize to one or more of the sequences in [SEQ ID NO:1] to [SEQ ID NO:14] or its complementary sequence. Such nucleic acid molecules preferably hybridize under high stringency conditions (see Sambrook et al. Molecular Cloning: A Laboratory Manual, Most Recent Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.). High stringency washes have preferably have low salt (preferably about 0.2% SSC) and a temperature of about 50-65 °C.

### Vaccines

One skilled in the art knows the preparation of live recombinant vaccines. Typically, such vaccines are prepared as injectables, either as liquid solutions or suspensions; solid forms suitable for solution in, or suspension in, liquid prior to injection may also be prepared. The preparation may also be emulsified, or the protein encapsulated in liposomes. The live immunogenic ingredients are often mixed with excipients that are pharmaceutically acceptable and compatible with the active ingredient. Suitable excipients are, for example, water, saline, dextrose, glycerol, ethanol, or the like and combinations thereof. In addition, if desired, the vaccine may contain minor amounts of auxiliary substances such as wetting or emulsifying agents, pH buffering agents, and/or adjuvants that enhance the effectiveness of the vaccine. Examples of adjuvants which may be effective include but are not limited to: aluminum hydroxide, N-acetyl-muramyl-L-threonyl-Disoglutamine (thr-MDP), N-acetyl-nor-muramyl-L-alanyl-D-isoglutamine (CGP 11637, referred to as nor-MDP), N-acetylmuramyl-L-alanyl-D-isoglutaminyl-L-alanine-2-(1'-2'dipalmitoyl-sn -glycero-3-hydroxyphosphoryloxy)-ethylamine (CGP 19835A, referred to as MTP-PE), and RIBI, which contains three components extracted from bacteria, monophosphoryl lipid A, trehalose dimycolate and cell wall skeleton (MPL+TDM+CWS) in a 2% squalene/Tween 80™ emulsion.

The effectiveness of an adjuvant may be determined by measuring the amount of antibodies directed against an immunogenic polypeptide containing a *Mycobacterium tuberculosis* antigenic sequence resulting from administration of the live recombinant *Mycobacterium bovis*-BCG vaccines that are also comprised of the various adjuvants. The vaccines are conventionally administered parenterally, by injection, for example, either subcutaneously or intramuscularly. Additional formulations which are suitable for

other modes of administration include suppositories and, in some cases, oral formulations. For suppositories, traditional binders and carriers may include, for example, polyalkylene glycols or triglycerides; such suppositories may be formed from mixtures containing the active ingredient in the range of 0.5% to 10%, preferably 1%-2%. Oral formulations include such normally employed excipients as, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate, and the like. These compositions take the form of solutions, suspensions, tablets, pills, capsules, sustained release formulations or powders and contain 10%-95% of active ingredient, preferably 25%-70%.

The vaccines are administered in a manner compatible with the dosage formulation, and in such amount as will be prophylactically and/or therapeutically effective.

The vaccine may be given in a single dose schedule, or preferably in a multiple dose schedule. A multiple dose schedule is one in which a primary course of vaccination may be with 1-10 separate doses, followed by other doses given at subsequent time intervals required to maintain and or reinforce the immune response, for example, at 1-4 months for a second dose, and if needed, a subsequent dose(s) after several months. The dosage regimen will also, at least in part, be determined by the need of the individual and be dependent upon the judgment of the practitioner.

In addition, the live recombinant *Mycobacterium bovis*-BCG vaccine administered in conjunction with other immunoregulatory agents, for example, immune globulins. A subject of the present invention is also a multivalent vaccine formula comprising, as a mixture or to be mixed, a live recombinant *Mycobacterium bovis*-BCG vaccine as defined above with another vaccine, and in particular another recombinant live recombinant *Mycobacterium bovis*-BCG vaccine as defined above, these vaccines comprising different inserted sequences.

### Pharmaceutical compositions

The pharmaceutical compositions of this invention are used for the treatmentment or prophylaxis of a mammal against challenge by *Mycobacterium tuberculosis* or *Mycobacterium bovis*. The pharmaceutical compositions of this invention are also used to treat patients having degenerative diseases, disorders or abnormal physical states such as cancer.

The pharmaceutical compositions can be administerd to humans or animals by methods such as tablets, aerosol administration, intratracheal instillation and intravenous injection.

### Media Compositions

The media compositions of this invention for inhibiting the growth of *Mycobacterium bovis*-BCG comprise alanine or serine as the only nitrogen source. When alanine is the only nitrogen source it is present in an amount of at least 0.03mM and when serine is the only nitrogen source it is present in an amount of at least 0.03mM.

The media compositions may further contain carbon in an amount of about 1.35g/L to about 1.65g/L, preferably in an amount of at least 1.5g/L; iron in an amount of about 0.045g/L to about 0.055g/L, preferably in an amount of at least 0.05g/L; magnesium in an amount of about 0.45g/L to about 0.55g/L, preferably in an amount of at least 0.5g/L; and SO<sub>4</sub> in an amount of about 0.045g/L to about 0.055g/L, preferably in an amount of at least 0.05g/L.

### **Kits**

Kits suitable for immunodiagnosis and containing the appropriate labeled reagents are constructed by packaging the appropriate materials, including the live recombinant *Mycobacterium bovis*-BCG strains of the instant invention, in suitable containers, along with the remaining reagents and materials required for the conduct of the assay, as well as a suitable set of assay instructions. Any immunological test format is contemplated, such as ELISA, Western blot, sandwich assay etc., which are well known to those skilled in the art.

### Materials and Methods

Bacterial strains and culture conditions. Twelve M. bovis BCG strains: BCG-Japan, BCG-Russia, BCG-Moreau, BCG-Sweden, BCG-Birkhaug BCG-Frappier, BCG-Pasteur, BCG-Glaxo, BCG-Phipps, BCG-Tice, BCG-Denmark, and BCG-Prague were used in this study and were obtained from Dr. Marcel Behr (McGill University). The identities of these strains were described in detail previously (Behr et al., 1999). Middlebrook 7H9 medium (Difco) contains (per liter) ammonium sulfate, 0.5 g; L-glutamate, 0.5 g; sodium citrate 0.1 g; pyridoxine, 1 mg; biotin, 0.5 mg; disodium phosphate 2.5g; monopotassium phosphate, 1 g; ferric ammonium citrate 40 mg; magnesium sulfate 50 mg; calcium chloride 0.5 mg; zinc sulfate 1 mg; copper sulfate, 1 mg; and glycerol, 2 ml; with 5 g of albumin (fraction V; bovine), 2 g of dextrose, and 0.05% Tween 80 added after sterilization. Sauton medium contains (per liter) L-asparagine, 4 g; monopotassium sulfate, 0.5 g; magnesium sulfate 0.5 g; ferric ammonium citrate 50 mg; citric acid, 2 g; zinc sulfate, 1 mg; and glycerol, 60 ml; with 0.05% Tween 80 added after sterilization. Glycerol-alanine-salts (GAS) medium contains (per liter) 2 g of ammonium chloride, 1 g of L-alanine, 0.3 g of Bacto Casitone (Difco), 4 g of dibasic potassium phosphate, 2 g of citric acid, 50 mg of ferric ammonium citrate, 1.2 g of magnesium chloride hexahydrate, 0.6 g of potassium sulfate, 1.8 ml of 10 M sodium hydroxide, and 10 ml of glycerol. Tween 80 was added to 0.05% after sterilization. BCG cultures were grown at 37°C with constant shaking for 3-4 weeks.

Cloning of ald. Cloning of ald [SEQ ID NO:1] was accomplished in two steps (Fig. 1). First, a 4.5kb Scal fragment of M. tuberculosis genomic DNA containing ald was ligated to Ecl136II-linearized pUC19 to generate pUC-ALD. Then mycobacterial plasmid pALD was created by ligating the 1.9 kb KpnI fragment containing the ald gene [SEQ ID NO:1] to KpnI- linearized pMD31 (Yu et al., 1998). The plasmid pALD was introduced by electroporation into M. bovis BCG, and recombinant M. bovis BCG selected on Middlebrook 7H9 agar (Difco) supplemented with 10% oleic/albumin/dextrose/catalase (OADC) enrichment and 25 µg/ml kanamycin.

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Cloning of sdaA. Cloning of sdaA [SEQ ID NO:5] was accomplished in two steps. First, a 9.5 kb BamHI fragment of M. tuberculosis genomic DNA was ligated to BamHI-linearized pMD31 to generate pSDA1. Plasmid pSDAA was generated by cleavage of pSDA1 with PstI, followed by self-ligation of the 10.9 kb PstI fragment. The plasmid pSDAA was introduced by electroporation into M. bovis BCG, and recombinant M. bovis BCG selected on Middlebrook 7H9 agar (Difco) supplemented with 10% oleic/albumin/dextrose/catalase (OADC) enrichment and 25 µg/ml kanamycin.

### Example 1

Growth of BCG strains in Glycerol-Alanine-Salts (GAS) medium. During the course of our studies, we found that BCG-Japan strain was able to grow in GAS medium, albeit slower than in 7H9 medium. BCG-Frappier and BCG-Pasteur could not grow in GAS medium, even after prolonged incubation (2 months). The growth of other BCG strains in GAS medium was also examined. The results are summarized in Table I, and show that BCG-Japan, BCG-Russia, BCG-Moreau, BCG-Sweden and BCG-Birkhaug were able to grow in GAS medium while BCG-Frappier, BCG-Pasteur, BCG-Glaxo, BCG-Phipps, BCG-Tice, BCG-Denmark, and BCG-Prague could not. This is an interesting observation since all 12 BCG strains listed above were able to grow in 7H9 and Sauton broth medium (Table I). To find out why certain BCG strains were unable to grow in GAS medium, the chemical compositions of GAS, 7H9 and Sauton medium were compared. Supplementing ZnSO<sub>4</sub> (1 mg/liter), which is present in 7H9 and Sauton but not in GAS medium, or sodium pyruvate (0.5%), which is required for growth of large colonies of M. bovis, did not support the growth of BCG strains in GAS (data not shown). Next, nitrogen sources were compared. L-Asparagine (4 g/liter) is the only nitrogen source in Sauton medium while ammonium chloride (2 g/liter) and L-alanine (1 g/liter) are the main nitrogen sources in GAS. When L-asparagine (at 4 g per liter) was added to GAS medium, BCG-Frappier, BCG-Pasteur, BCG-Glaxo, BCG-Phipps, BCG-Tice, BCG-Denmark, and BCG-Prague were able to grow rapidly (Table I). Supplementing L-aspartate, Lglutamine, or L-glutamate but not other types of amino acids to GAS medium also supported the growth of these BCG strains (Table I). These results show that the failure

of certain BCG strains to grow in GAS medium is caused by their inability to utilize the nitrogen source present.

### Example 2

Amino acids as the nitrogen source for growth of BCG strains. The above result prompted us to examine the ability of BCG strains to utilize various types of amino acids as the only nitrogen source. Since GAS medium contains a small amount of Bacto Casitone (0.3 g/liter), which is a complex mixture of various amino acids and peptides, we chose Sauton medium, which is a defined medium, for this purpose. The L-asparagine in the original formula for Sauton medium was replaced individually by each type of amino acids at the same concentration (27 mM), and pH was adjusted to 7.0. Ammonium chloride at 27 mM or 1 mM as the only nitrogen source was also tested. Table II summarizes the results for three representative BCG strains, BCG-Japan, BCG-Pasteur, and BCG-Frappier. Consistent with the result in Table I, all three BCG strains grew rapidly when L-asparagine, L-aspartate, L-glutamine, or L-glutamate was used as the only nitrogen source. BCG-Japan was able to grow on cationic amino acids (e.g., L-arginine, L-lysine) while BCG-Pasteur and BCG-Frappier could not. More interestingly, none of the BCG strains were able to utilize L-alanine, L-serine, L-leucine, L-isoleucine, Lmethioine, or L-glycine as the only nitrogen source, while other Mycobacterium species, including pathogenic M. tuberculosis and M. avium, and nonpathogenic M. smegmatis, were able grow on these amino acids. These results demonstrate that BCG vaccine strains utilize limited types of amino acids as the nitrogen source for growth; some BCG strains such as BCG-Pasteur or BCG-Frappier can grow only on 4 types of amino acids (Table II). Such a limitation is likely to restrict the ability of BCG to grow and persist in vivo (within the host).

### Example 3

L-Alanine, D-alanine, or L-serine inhibits the growth of BCG. One surprising finding from the above experiment was that all BCG strains are able to grow on ammonium chloride as the only nitrogen source at both low (1 mM) or high concentrations (27 mM) (Table II). This is contradictory to the result obtained in GAS medium, in which

ammonium chloride at 37 mM does not support the growth of BCG-Pasteur and BCG-Frappier (Table I). Since GAS medium also contains L-alanine, and L-alanine is not utilized by BCG strains for growth (Table II), the only possible explanation is that L-alanine actually inhibits the growth of BCG strains. To prove this, a modified GAS medium, in which L-alanine was omitted, was made and the growth of BCG strains in this medium was examined. As predicted, BCG-Frappier and BCG-Pasteur, which are unable to grow in the original GAS medium containing L-alanine, grew rapidly in GAS without L-alanine (Fig. 3). BCG-Japan also grew more rapidly in this L-alanine free medium than in the original GAS medium (Fig. 3). The same results were obtained for the other nine BCG strains listed in this report.

To further confirm this result, increasing concentrations of L-alanine were added to Sauton medium containing ammonium chloride (5 g/liter) and the growth of BCG-Japan, BCG-Frappier and BCG-Pasteur was determined (Fig. 4). Strikingly, even at a very low concentration (0.25 mM), L-alanine completely inhibited the growth of BCG-Frappier and BCG-Pasteur. Although the growth inhibition of BCG-Japan was somewhat less severe, L-alanine at 0.5 mM significantly reduced its growth and at 8-16 mM the growth was completely inhibited (Fig. 4). Taken together, these results clearly demonstrate that L-alanine inhibits the growth of BCG strains. We further found that D-alanine also inhibits the growth of BCG strains. The presence of D-alanine in GAS medium stopped the growth of BCG-Pasteur and BCG-Frappier, and significantly reduced the growth of BCG-Japan (Fig. 5). Similarly, the presence of L-serine in GAS medium significantly inhibited the growth of BCG-Japan, BCG-Frappier, and BCG-Pasteur (Fig. 7).

### Example 4

Expressing L-alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2] in BCG relieves the inhibition of BCG growth by L-alanine and D-alanine. Alanine is an excellent source of nitrogen for many *Mycobacterium* species including *M. tuberculosis*, *M. avium*, and *M. smegmatis*. D-Alanine degradation begins with racemization to L-alanine, which is then broken down to ammonium and pyruvate by L-alanine dehydrogenase. Interestingly, a functional L-alanine dehydrogenase was detected in *M*.

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tuberculosis and M. smegmatis but not in BCG-Japan or BCG-Copenhagen (Andersen et al., 1992; Hutter and Dick, 1998). We did not detect L-alanine dehydrogenase activity in any of the BCG strains listed in this study (data not shown). The failure of BCG strains to utilize L- or D- alanine as the only nitrogen source for growth is due to the lack of a functional L-alanine dehydrogenase. To prove this, the ald gene [SEQ ID NO:1] coding for L-alanine dehydrogenase [SEQ ID NO:2] in the M. tuberculosis genome was cloned into a shuttle vector and transformed into BCG-Frappier and BCG-Pasteur. The resulting recombinant BCG strains were tested for their ability to grow in GAS medium containing L-alanine or D-alanine. Both recombinant strains, BCG-Frappier/ald and BCG-Pasteur/ald, grew rapidly in GAS medium containing either L-alanine or D-alanine (Fig. 6), while strains containing the cloning vector alone did not grow. This result shows that expression of a functional L-alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2] in BCG strains relieves the growth inhibition of BCG by L-alanine and D-alanine.

### Example 5

Expressing L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6] in BCG relieves the inhibition of BCG growth by L-serine. L-Serine is used by M. tuberculosis, M. avium and M. smegmatis, but not M. bovis BCG, as the only nitrogen for growth. The failure of BCG to utilize L-serine as the only nitrogen source is likely to be caused by either mutations on or altered expression of the gene encoding L-serine dehydratase, sdaA [SEQ ID NO:5], in BCG. Expression of sdaA [SEQ ID NO:5; SEQ ID NO:6] of M. tuberculosis in BCG allows BCG strains to grow on L-serine as the only nitrogen source and relieves the inhibition of BCG growth by L-serine (Fig. 8).

### Example 6

Inhibition of BCG growth by L-alanine, D-alanine and L-serine are likely to occur by blocking the activity of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO:14]. Glutamine synthetase plays a central role in nitrogen metabolism in bacteria (Reitzer, 1996). Working in tandem with glutamate synthase, glutamine synthetase catalyzes the synthesis of glutamine and glutamate, which together provide nitrogen for almost all amino acids, proteins, and nucleotides. In *Escherichia coli* and *Klebsiella aerogenes*,

glutamine synthetase is under feedback inhibition - purified glutamine synthetase is inhibited by L-alanine, L-serine and glycine (Reitzer, 1996). Glutamine synthetase was identified as an extracellular protein in M. tuberculosis and M. bovis BCG (Harth et al., 1994). It is likely that undegraded L-alanine inhibits glutamine synthetase and subsequently prevents the growth of BCG. If this were correct, then L-serine, which was not catabolized by BCG for growth (Table I), would also inhibit the growth of BCG by the same mechanism. Supporting this hypothesis, addition of L-serine to GAS medium containing only ammonium chloride as the nitrogen source inhibits the growth of BCG-Frappier, BCG-Pasteur or BCG-Japan (Fig. 7). Furthermore, if glutamine synthetase were the target of L-alanine and L-serine inhibition, then supplementing amino acids that can be converted to glutamate would also alleviate their effects, as demonstrated in K. aerogenes (Janes and Bender, 1998). Indeed, addition of L-glutamate and amino acids that could be catabolized to yield glutamate (L-glutamine, L-asparagine, and L-aspartate) allows the growth of BCG strains in the presence of alanine (Table I), but those that could not be catabolized to glutamate (e.g., L-lysine, L-methioine, L-leucine) fail to allow growth. BCG-Frappier and BCG-Pasteur are more sensitive than BCG-Japan to inhibition by alanine and serine, this is due to differences in the expression level or activity of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO:14], i.e., BCG-Japan produces more glutamine synthetase or with higher activity than BCG-Frappier or BCG-Pasteur.

The present invention has been described in detail and with particular reference to the preferred embodiments; however, it will be understood by one having ordinary skill in the art that changes can be made without departing from the spirit and scope thereof. For example, where the application refers to proteins, it is clear that peptides and polypeptides may often be used. Likewise, where a gene is described in the application, it is clear that nucleic acids or gene fragments may often be used.

All publications (including Genbank entries), patents and patent applications are incorporated by reference in their entirety to the same extent as if each individual publication, patent or patent application was specifically and individually indicated to be incorporated by reference in its entirety.

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Table I

Comparative growth of M. tuberculosis, M. smegmatis and M. bovis BCG substrains in 7H9, Sauton, and glycerol-alanine-salts (GAS) medium.

	(H)	Sauton	GAS				
M tuherculosis <sup>6</sup>	+	+	+	+	+	+	+
M. smegmatis	+	+	+	+	+	+	+
BCG-Russia	+	+	+	+	+	+	+
RCC-Moreau	+	+	+	+	+	+	+
RCG-Janan	+	+	+	+	+	+	+
BCC Sweden	+	+	+	+	+	+	+
	+	+	+	+	+	÷	+
BCG-birkusug	· +	+	•	+	+	+	+
BCG-Frague	+	+	•	+	+	+	+
BCG-Denmark	+	+	•	+	+	+	· +

				4	+	+	+
BCG-Tice	+	+	1	٠			
BCG-Frappier	+	+	•	+	+	+	+
•	_	+	•	+	+	+	+
BCG-Phipps	ŀ	•				-1	+
BCG-Pasteur	+	+	1	+	+	-	

<sup>a</sup> Each 5 ml culture inoculated with  $1\times10^7$  cells of M. smegmatis or M. bovis BCG substrains.

<sup>b</sup> L-Asn, L-Asp, L-Glu and L-Gln in GAS supplemented to a final concentration of 27 mM.

° Based on research literature.

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Table II

Comparative growth of M. bovis BCG-Japan, BCG-Frappier, BCG-Pasteur, M. tuberculosis, M. avium and M. smegmatis

Media	BCG-Japan	BCG-Frappier <sup>b</sup>	BCG-Pasteur <sup>b</sup>	M. tuberculosis°	M. avium <sup>c</sup>	M. smegmatis
Sauton basal	,	1	1	1		•
Group 1						
Sauton + L-Asn	++++	+++	++++	++++	+ + +	+ + +
Santon + 1. Asp	+ + +	+ + +	++++	+ + +	+ + +	+ + +
Souton +1.Clu	+ + +	+ + +	+ + +	+ + +	+ + +	+ + +
HD 17 money	++++	+ + +	+ + +	+ + +	+ + +	+ + +
	+++++++++++++++++++++++++++++++++++++++	+ + +	++	+ + +	++++	++++
Sauton + L-Cys Sauton + NH.Cl	+ + +	+ + +	+ + +	+ + +	+ + +	++++
Groun 2						
r droip	-	•	,	++++	+++	++++
Sauton + L-Arg	+ +				-	+++++++++++++++++++++++++++++++++++++++
Sauton + L-His	+++	•	ŧ	+ + +	<del> </del> <del> -</del>  -	-

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						447
Sauton + L-Lys	++		•	NA	+ + +	- -
Souten + 1 -Pro	+ +		ı	NA	1	+ + +
Sauton + CARA	<del>+</del> +	ı	ı	NA	NA	+ + +
Sauton + L-Ornithine	+ +	ı	•	NA	NA	+ + +
, 1						
Group 3				+++++++++++++++++++++++++++++++++++++++	++++	+++++
Sauton + L-Ala	•	•	1		+ + +	+ + +
Sauton + L-Ser	ı			+ + +	+ + +	
Sauton +1~Leu	•	•		+ + +	+ + +	+ + +
	•	•	,	+ + +	+ + +	+ + +
Sauton + L-11e				ĄN	+ + +	+ + +
Sauton + L-Met	1	•	•		;	<b>4</b> 4 4
Sauton + Glycine	•	•	•	+ + +	NA	-
Group 4						
					,	1
Sauton + L-Trp	3	•	•			1
Sauton + L.Phe	t	•		+ + +	2	,

	1		
•	•	•	
•	NA	NA	
	r	•	
•	•	,	
	,	t	
Sauton + L-Tyr	Sauton + L-Val	Sauton + L-Thr	

<sup>2</sup> All amino acids, L-Ornithine and GABA supplemented to final concentration of 27mM. NH4Cl was tested at 1mM, 27 mM and 96 mM.

 $^{b}$  Each 5 ml culture inoculated with  $1\times10^{7}$  cells of M. smegmatis or M. bovis BCG substrains.

<sup>c</sup> Based on research literature.

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#### We claim:

- 1. A live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid apable of expression, the nucleic acid encoding at least one protein or polypeptide that exhibits alanine dehydrogenase activity [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase activity [SEQ ID NO:7 to SEQ ID NO:14], or L-serine dehydratase activity [SEQ ID NO:5; SEQ ID NO:6].
- 2. A live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide selected from the group consisting of alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase [SEQ ID NO:7 to SEQ ID NO:14] and L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6].
- 3. A live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid comprises all or part of at least one nucleic acid molecule selected from the group consisting of [SEQ ID NO:1], [SEQ ID NO:2], [SEQ ID NO:5], [SEQ ID NO:6], [SEQ ID NO:7], [SEQ ID NO:8], [SEQ ID NO:9], [SEQ ID NO:10], [SEQ ID NO:11], [SEQ ID NO:12], [SEQ ID NO:13], and [SEQ ID NO:14].
- 4. The live recombinant Mycobacterium bovis-BCG strain of claim 1, 2 or 3 wherein the Mycobacterium bovis-BCG strain is selected from the group consisting of Mycobacterium bovis-BCG-Russia, Mycobacterium bovis-BCG-Moreau, Mycobacterium bovis-BCG-Japan, Mycobacterium bovis-BCG-Sweden, Mycobacterium bovis-BCG-Birkhaug, Mycobacterium bovis-BCG-Prague, Mycobacterium bovis-BCG-Glaxo, Mycobacterium bovis-BCG-Denmark, Mycobacterium bovis-BCG-Tice, Mycobacterium bovis-BCG-Frappier, Mycobacterium bovis-BCG-Connaught, Mycobacterium bovis-BCG-Phipps, and Mycobacterium bovis-BCG-Pasteur.
- 5. A pharmaceutical composition comprising the live recombinant *Mycobacterium bovis*-BCG strain of claim 1, 2 or 3.

- 6. A vaccine or immunogenic composition for treatment or prophylaxis of a mammal against challenge by mycobacteria comprising the live recombinant *Mycobacterium bovis*-BCG strain of claim 1, 2 or 3.
- 7. The vaccine or immunogenic composition of claim 6 wherein the mycobacteria is *Mycobacterium tuberculosis*.
- 8. The vaccine or immunogenic composition of claim 6 or 7 further comprising a pharmaceutically acceptable carrier.
- 9. The vaccine or immunogenic composition of claim 6, 7, or 8 further comprising an adjuvant.
- 10. The vaccine or immunogenic composition of claim 6, 7, 8 or 9 further comprising immunogenic materials from one or more other pathogens.
- 11. A method for treatment or prophylaxis of a mammal against challenge by *Mycobacterium tuberculosis* or *Mycobacterium bovis* comprising administering to the mammal the vaccine or immunogenic composition of claim 1, 2 or 3.
- 12. The method of claim 11 wherein the mammal is a cow.
- 13. The method of claim 11 wherein the mammal is a human.
- 14. The method of claim 11 wherein the vaccine or immunogenic composition is administered in the presence of an adjuvant.
- 15. A method for treatment or prophylaxis of a mammal against cancer comprising administering to the mammal the vaccine or immunogenic composition of claim 1, 2 or 3.
- 16. The method of claim 15 wherein the vaccine or immunogenic composition is administered in the presence of an adjuvant.
- 17. The method of claim 15 or 16 wherein the cancer is bladder cancer.

- 18. A test kit comprising the live recombinant Mycobacterium bovis-BCG strain of claim 1, 2 or 3.
- 19. A media composition for inhibiting the growth of *Mycobacterium bovis-BCG* comprising alanine as the only nitrogen source for growth.
- 20. A media composition for inhibiting the growth of *Mycobacterium bovis-BCG* comprising serine as the only nitrogen source for growth.
- 21. The media composition of claim 19 or 20 further comprising:
- (a) a carbon source;
- (b) iron;
- (c) magnesium; and
- (d) SO<sub>4</sub>.
- 22. A media composition of claim 21 wherein the carbon source is selected from the group consisting of glycerol, dextrose, citrate and glucose.
- 23. A method for inhibiting the growth of Mycobacterium bovis-BCG comprising:
- (a) obtaining a sample comprising Mycobacterium; and
- (b) culturing the sample in a selective media.
- 24. The method of claim 23, wherein the selective media comprises alanine as the only nitrogen source for growth.
- 25. The method of claim 23, wherein the selective media comprises serine as the only nitrogen source for growth.
- 26. A method of culturing Mycobacterium bovis-BCG comprising:
- (a) obtaining a sample of Mycobacterium; and

- (b) culturing the sample in differential media.
- 27. The method of claim 26, wherein the differential media comprises histidine.

## Abstract

The invention relates to a live recombinant Mycobacterium bovis-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide that exhibits alanine dehydrogenase activity, glutamine synthetase activity, or serine dehydratase activity.

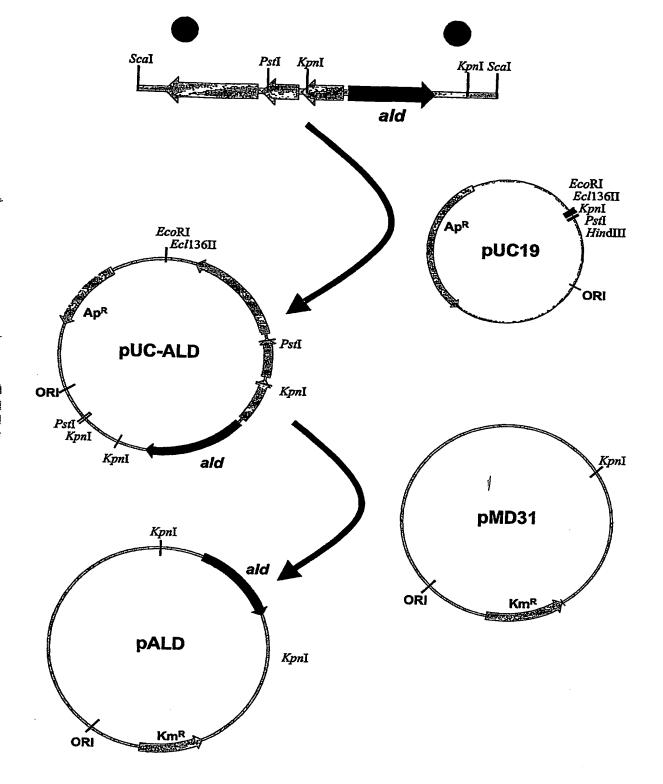


Fig. 1

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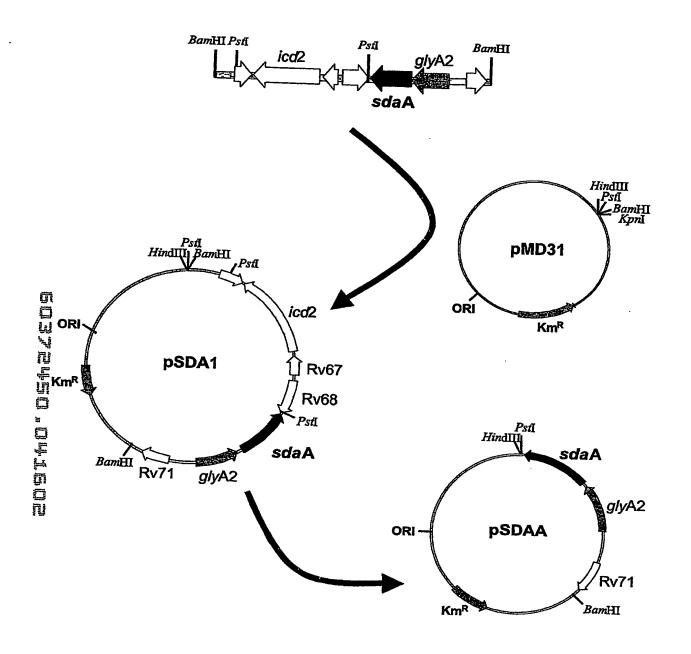


Fig. 2

Conversed L. Hora

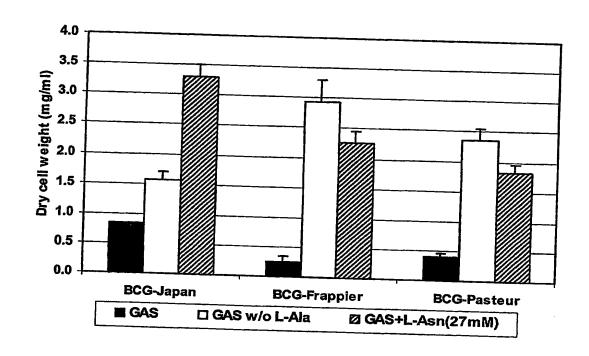


Fig. 3

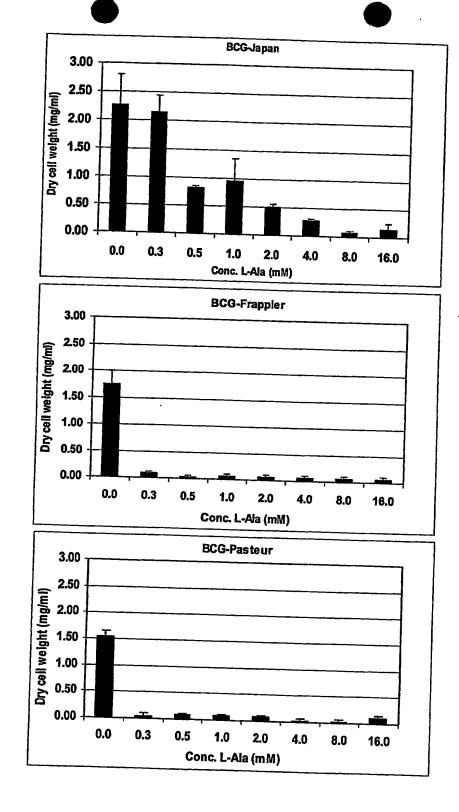


Fig. 4

Conversalited to trans-

a)

b)

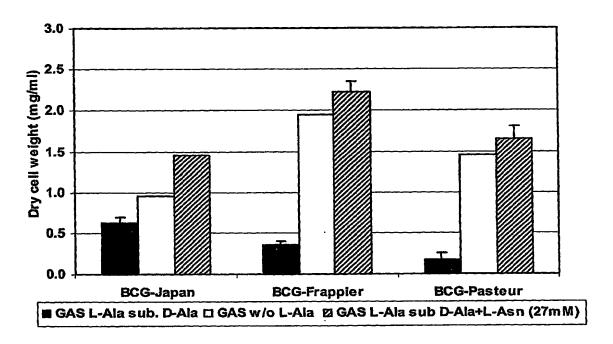


Fig. 5

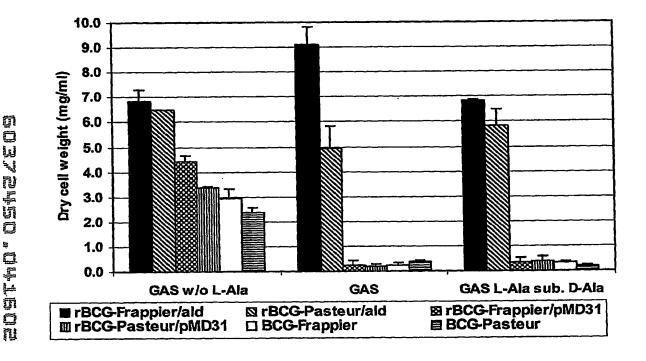
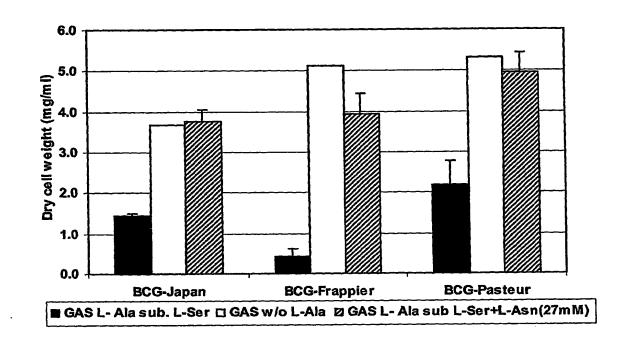


Fig. 6



**Fig. 7** 

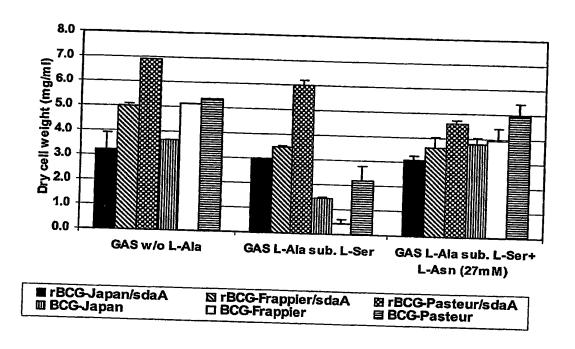


Fig. 8

B

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M.tb	ACC CCG															
M.bovis																
M.tb M.bovis	GCA GGT GCA GGT															
M.tb M.bovis	GCG CAA															
M.tb M.bovis	AAG GTC AAG GTC															
_												200	<b></b>	000	mma	mma
M.tb M.bovis	TTG TTC TGT TCA															
M.tb M.bovis	GAT TCC ATT CCG															
M.tb M.bovis	CTA CCC					AGC	gaa	GTC	GCC	GGT	CGA	CTC	GCC	GCC	CAG	GTT
M. tb	GGC GCT	TAC CA	CTG	atg	CGA	ACC	CAA	GGG	GGC	CGC	GGT	GTG	CTG	ATG	GGC	GGG
M. tb	GTG CCC	GGC GT	GAA	CCG	GCC	GAC	GTC	GTG	GTG	ATC	GGC	GCC	GGC	ACC	GCC	GGC
M. tb	TAC AAC	GCA GC	c ccc	ATC	GCC	AAC	GGC	ATG	GGC	GCG	ACC	GTT	ACG	GTT	CTA	GAC
. M.tb	ATC AAC	ATC GA	C AAA	CTT	CGG	CAA	CTC	GAC	GCC	GAG	TTC	TGC	GGC	CGG	ATC	CAC
M. tb	ACT CGC	TAC TC	A TCG	GCC	TAC	GAG	CTC	GAG	GGT	GCC	GTC	AAA	CGT	GCC	GAC	CIG
M. tb	GTG ATT	GGG GC	C GTC	CTG	GTG	CCA	GGC	GCC	AAG	GCA	ccc	AAA	TTA	GTC	TCG	AAT
M. tb	TCA CTT	GTC GC	G CAT	ATG	AAA	CCA	GGT	GCG	GTA	CTG	GTG	GAT	ATA	GCC	ATC	GAC
M. tb	CAG GGC	GGC TG	r TTC	gaa	GGC	TCA	CGA	CCG	ACC	: ACC	TAC	GAC	CAC	cca	ACG	TTC
M. tb	GCC GTG	CAC GA	C ACG	CTG	TTT	TAC	TGC	GTG	GCG	AAC	ATG	ccc	GCC	TCG	GTG	CCC
M. tb	aag acg	TCG AC	C TAC	: GCG	CTG	ACC	AAC	GCG	ACC	ATG	CCG	TAT	GTG	CTC	GAG	CTT
M. tb	GCC GAC	CAT GG	C TGG	CGG	GCG	GCG	TGC	CGG	TCC	TAA :	. cca	GCA	CTA	GCC	AAA	GGT
M. tb	CTT TCG	acg ca	C GAA	GGG	GCG	TTA	CTG	TCC	GAI	CGG	GTG	GCC	ACC	: GAC	CTG	GGG
M.Eb M.bovis	GTG CCG MRVGIPT MRVGIPT															
M.tb M.bovis	LKVKEPI LKVKEPI														SEVAC	RLAA
M. tb	QVGAYHL	MRTQGGR	GVLMC	GVPG	VEPA	יסטענ	/IGAG	'EDAT	IAAN	NAIS	MGAT	VIVI	DINI	DKL	RQLDI	ÆFCG
M. tb	RIHTRYS	SAYELEG	AVKR	DLAI	GAVI	'Abgi	KAPI	CLVS	ISLVI	AHMKI	GAVI	'ADI'	DQC	GCF1	ZGSRI	PTTYD
M. tb	HPTFAVH	DTLFYCV	'ANMP	4SA51	TST	ALT	IATMI	YVLE	ELADI	HGWRI	ACRE	NPAI	AKGI	LSTHI	EGALI	SERV
M. tb	ATDLGVP	FTEPASV	LA-													

Fig. 9

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Leu Glu Gly Ala Val Lys Arg Ala Asp Leu Val Ile Gly Ala Val Leu 225 230 235 240

Val Pro Gly Ala Lys Ala Pro Lys Leu Val Ser Asn Ser Leu Val Ala 245 250 255

His Met Lys Pro Gly Ala Val Leu Val Asp Ile Ala Ile Asp Gln Gly 260 265 270

Gly Cys Phe Glu Gly Ser Arg Pro Thr Thr Tyr Asp His Pro Thr Phe 275 280 285

Ala Val His Asp Thr Leu Phe Tyr Cys Val Ala Asn Met Pro Ala Ser 290 295 300

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Trp 65	Ala	Asp	Ala	Asp	Leu 70	Leu	Leu	ГÀв	Val	<b>Lу</b> в 75	Glu	Pro	Ile	Ala	Ala 80	
Glu	туг	Gly	Arg	Leu 85	Arg	His	Gly	Arg	Ser 90	Сув	Ser	Arg	Ser	Суз 95	Ile	
Trp	Pro	Arg	His 100	Val	Leu	Ala	Pro	Met 105	Arg	Сув	Trp	Ile	Pro 110	Ala	Pro	
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		Segu	ence	is		tica	1 to	the	com			of n	ucle	otid	es 1319!	5-14580
atg		atc													tcc Ser	48
agt Ser	tcc Ser	cac His	acc Thr 20	gtg Val	gga Gly	ccg	atg Met	cgc Arg 25	gcg Ala	gca Ala	aac Asr	cag Glr	ttc Phe 30	gta Val	gtt Val	96
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A	la	Leu	Arg 35	Arg	Arg	Gly	His	Leu 40	Asp	Двр	Leu	Glu	Ala 45	Met	Arg	Val		
g A	at sp	ctg Leu 50	ttc Phe	ggc Gly	tcg Ser	ctc Leu	gcg Ala 55	gcc Ala	acc Thr	gga Gly	gcc Ala	ggt Gly 60	cat His	ggc Gly	acc Thr	atg Met	1	L92
S	cg er 5	gcg Ala	ata Ile	ttg Leu	ctg Leu	999 Gly 70	ctg Leu	gaa Glu	Gly ggc	tgc Cys	cag Gln 75	cca Pro	gaa Glu	acg Thr	att Ile	acc Thr 80	2	240
a T	cc hr	gaa Glu	cac His	aag Lys	gaa Glu 85	cgc Arg	cgg Arg	ctc Leu	gcc Ala	gag Glu 90	atc Ile	gca Ala	gcg Ala	tcc Ser	ggc Gly 95	gtg Val	;	288
a T	cg hr	cga Arg	atc Ile	ggc Gly 100	ggt Gly	gtc Val	att Ile	ccg Pro	gtc Val 105	ccg Pro	ctg Leu	acc Thr	gag Glu	cgt Arg 110	gat Asp	atc Ile	:	336
g A	ac sp	ctg Leu	cat His 115	ccc Pro	gac Asp	atc Ile	gtt Val	ctg Leu 120	cca Pro	acg Thr	cat His	ccc Pro	aac Asn 125	gga Gly	atg Met	acg Thr	;	384
						cca Pro											,	432
P	tt he 45	tcg Ser	gtg Val	Gly	gga Gly	999 91y	ttc Phe	atc Ile	gtc Val	acg Thr	gaa Glu 155	cag Gln	acc Thr	agc Ser	ggc	aac Asn 160		480
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9	gaa Blu	ctg Leu	ctg Leu	gac Asp 180	Ile	tgt Cys	gac Asp	cgc Arg	ctc Leu 185	qaA	gtg Val	tca Ser	att Ile	agc Ser 190	Glu	gcg Ala		576
Ş	geg Ma	ctg Leu	cgc Arg 195	Asn	gaa Glu	aca Thr	tgt Cys	tgc Cys 200	Arg	acc Thr	gag	aac Asn	gag Glu 205	Val	cgc Arg	gcc		624
Ş	ycg Lla	ctg Leu 210	Leu	cac His	ctg Leu	cgc Arg	gac Asp 215	Val	atg Met	gtt Val	gag Glu	tgo Cys 220	Glu	cag Gln	cgg Arg	agc Ser		672
]		Āla										Arg				cga Arg 240		720
2	gcg Ala	aag Lys	gtg Val	tgg Trp	tat Tyr 245	Asp	cgc	ttg Lev	aac Asn	gco Ala 250	Glu	gac Asp	ccc Pro	act Thr	cgc Arg 255	aag Lys		768
3	ccg Pro	gaa	ttc Phe	gct Ala	gag Glu	gac Asp	tgg Trp	gto Val	aac . Asr	cto	gto Val	gcg Ala	g cto	g gca	gto Val	aac		816

270 260 265 gag gag aac gcc tec ggt ggg cgc gtc gtc acc gcc ccg acc aac ggt 864 Glu Glu Asn Ala Ser Gly Gly Arg Val Val Thr Ala Pro Thr Asn Gly 280 912 gcc gcc ggc atc gtg ccg gcg gtc ctg cac tac gca atc cac tac acg Ala Ala Gly Ile Val Pro Ala Val Leu His Tyr Ala Ile His Tyr Thr tcg gcc ggc gcg ggg gac ccc gac gat gtc acc gtg cga ttc ctg ctc 960 Ser Ala Gly Ala Gly Asp Pro Asp Asp Val Thr Val Arg Phe Leu Leu act gct gga gcc atc gga tcg ttg ttc aag gag cga gca tcg atc tcc 1008 Thr Ala Gly Ala Ile Gly Ser Leu Phe Lys Glu Arg Ala Ser Ile Ser 330 1056 gga gcc gag gtc ggc tgt cag ggc gag gtc ggc tcc gcg gcc gcc atg Gly Ala Glu Val Gly Cys Gln Gly Glu Val Gly Ser Ala Ala Ala Met gcc gcc gcc gga ttg gct gaa atc ctc ggc ggc aca ccg cga caa gtg 1104 Ala Ala Ala Gly Leu Ala Glu Ile Leu Gly Gly Thr Pro Arg Gln Val 360 1152 gaa aac gcc gcc gag atc gcc atg gaa cac agc ctc ggc ctg acc tgt Glu Asn Ala Ala Glu Ile Ala Met Glu His Ser Leu Gly Leu Thr Cys 375 1200 gac ecc ate gee ggg etg gtg cag ate ecc tge ate gaa ege aac geg Asp Pro Ile Ala Gly Leu Val Gln Ile Pro Cys Ile Glu Arg Asn Ala 390 395 1248 att tee gee gge aag gee ate aac gee gea egg atg gea ttg ege gge Ile Ser Ala Gly Lys Ala Ile Asn Ala Ala Arg Met Ala Leu Arg Gly 405

ggg ctc gcc atc aac gtc gca gtc aac atc gtc gag tgt tga Gly Leu Ala Ile Asn Val Ala Val Asn Ile Val Glu Cys 450 455 460

gac ggc atc cat cgc gtc acc ctc gac cag gtc atc gac acc atg cgc

Asp Gly Ile His Arg Val Thr Leu Asp Gln Val Ile Asp Thr Met Arg

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Ala Thr Gly Ala Asp Met His Thr Lys Tyr Lys Glu Thr Ser Ala Gly

1296

1344

1386

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<211> 461

435

<212> PRT

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<220>

<221>

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<223> Sequence is identical to SwissProt entry SP:SDHL\_MYCTU Sequence is identical to GenBank entries GP:AE006919\_13 and GP:MTV030\_11

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Ala Leu Arg Arg Gly His Leu Asp Asp Leu Glu Ala Met Arg Val 35 40 45

Asp Leu Phe Gly Ser Leu Ala Ala Thr Gly Ala Gly His Gly Thr Met 50 55 60

Ser Ala Ile Leu Leu Gly Leu Glu Gly Cys Gln Pro Glu Thr Ile Thr 65 70 75 80

Thr Glu His Lys Glu Arg Arg Leu Ala Glu Ile Ala Ala Ser Gly Val 85 90 95

Thr Arg Ile Gly Gly Val Ile Pro Val Pro Leu Thr Glu Arg Asp Ile
100 105 110

Asp Leu His Pro Asp Ile Val Leu Pro Thr His Pro Asn Gly Met Thr 115 120 125

Phe Thr Ala Ala Gly Pro His Gly Arg Val Leu Ala Thr Glu Thr Tyr 130 135 140

Phe Ser Val Gly Gly Phe Ile Val Thr Glu Gln Thr Ser Gly Asn 145 150 155 160

Ser Gly Gln His Pro Cys Ser Val Ala Leu Pro Tyr Val Ser Ala Gln
165 170 175

Glu Leu Leu Asp Ile Cys Asp Arg Leu Asp Val Ser Ile Ser Glu Ala 180 185 190

Ala Leu Arg Asn Glu Thr Cys Cys Arg Thr Glu Asn Glu Val Arg Ala 195 200 205 Ala Leu Leu His Leu Arg Asp Val Met Val Glu Cys Glu Gln Arg Ser 210 215 220

Ile Ala Arg Glu Gly Leu Leu Pro Gly Gly Leu Arg Val Arg Arg 225 230 235 240

Ala Lys Val Trp Tyr Asp Arg Leu Asn Ala Glu Asp Pro Thr Arg Lys 245 250 255

Pro Glu Phe Ala Glu Asp Trp Val Asn Leu Val Ala Leu Ala Val Asn 260 265 270

Glu Glu Asn Ala Ser Gly Gly Arg Val Val Thr Ala Pro Thr Asn Gly 275 280 285

Ala Ala Gly Ile Val Pro Ala Val Leu His Tyr Ala Ile His Tyr Thr 290 295 300

Ser Ala Gly Ala Gly Asp Pro Asp Asp Val Thr Val Arg Phe Leu Leu 305 310 315 320

Thr Ala Gly Ala Ile Gly Ser Leu Phe Lys Glu Arg Ala Ser Ile Ser 325 330 335

Gly Ala Glu Val Gly Cys Gln Gly Glu Val Gly Ser Ala Ala Ala Met 340 345 350

Ala Ala Gly Leu Ala Glu Ile Leu Gly Gly Thr Pro Arg Gln Val 355 360 365

Glu Asn Ala Ala Glu Ile Ala Met Glu His Ser Leu Gly Leu Thr Cys 370 375 380

Asp Pro Ile Ala Gly Leu Val Gln Ile Pro Cys Ile Glu Arg Asn Ala 385 390 395 400

Ile Ser Ala Gly Lys Ala Ile Asn Ala Ala Arg Met Ala Leu Arg Gly
405 410 415

Asp Gly Ile His Arg Val Thr Leu Asp Gln Val Ile Asp Thr Met Arg
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Ala Thr Gly Ala Asp Met His Thr Lys Tyr Lys Glu Thr Ser Ala Gly 435 440 445

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<212> DNA
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<220>
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<222> (1)..(1437)
<223> Sequence is identical to General Control 
<223> Sequence is identical to GenBank entry GB:MTU87280 [U87280] Sequence is identical to nucleotides 163-1599 of GenBank entry GB:MTCY427

[270692]

Sequence is identical to nucleotides 93-1529 of GenBank entry GB:AE007073

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														ggc Gly 30			96	
	_			_		_		_	_		-	-		gtg Val	_		144	
														cag Gln			192	
														gcg Ala			240	,
														ttt Phe			288	ļ
														aac Asn 110			336	;
•									Ser					Ąsp		gca Ala	384	Ļ
	tac	ttc	ggc	gcc	gag	gcc	gag	ttc	tac	att	ttc	gat	tcg	gtg	agc	ttc	432	2

Tyr	Phe 130	Gly	Ala	Glu	Ala	Glu 135	Phe	Tyr	Ile	Phe	Asp 140	Ser	Val	Ser	Phe	
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ggc	tac Tyr	aag Lys	gec Val 180	cgc Arg	cac His	aag Lys	ggc Gly	999 Gly 185	tat Tyr	ttc Phe	cca Pro	gtg Val	gcc Ala 190	ccc Pro	aac Asn	576
gac gac	caa Gln	tac Tyr 195	gtc Val	gac Asp	ctg Leu	cgc Arg	gac Asp 200	aag Lys	atg Met	ctg Leu	acc Thr	aac Asn 205	ctg Leu	atc Ile	aac Asn	624
tcc Ser	ggc Gly 210	ttc Phe	atc Ile	ctg Leu	gag Glu	aag Lys 215	ggc Gly	cac His	cac His	gag Glu	gtg Val 220	ggc Gly	agc Ser	ggc Gly	gga Gly	672
cag Gln 225	gcc Ala	gag Glu	atc Ile	aac Asn	tac Tyr 230	cag Gln	ttc Phe	aat Asn	tcg Ser	ctg Leu 235	ctg Leu	cac His	gcc Ala	gcc Ala	gac Asp 240	720
gac Asp	atg Met	cag Gln	ttg Leu	tac Tyr 245	aag Lys	tac Tyr	atc Ile	atc Ile	aag Lys 250	aac Asn	acc Thr	gcc Ala	tgg Trp	cag Gln 255	Asn	768
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atg Met	tac Tyr 290	Asp	gag Glu	acg Thr	ggt	tat Tyr 295	Ala	ggt	ctg Leu	tcg Ser	gac 300	Thr	gcc	cgt Arg	cat His	912
tac Tyr 305	Ile	ggg	ggc Gly	ctg Leu	tta Leu 310	His	cac His	gcg Ala	ccg Pro	tcg Ser 315	Leu	ctg Leu	gcc Ala	tto Phe	acc Thr 320	960
aac Asn	ccg Pro	acg Thr	gtg Val	aac Asn 325	Ser	tac Tyr	aag Lys	cgg Arg	ctg Leu 330	Val	ecc Pro	ggt Gly	tac Tyr	gag Glu 335	gcc Ala	1008
ccg Pro	atc Ile	aac Asr	ctg Leu 340	Val	tat Tyr	ago Ser	cag Glr	g cgc 1 Arg 345	J Asi	e egg	j te <u>e</u> j Ser	gca Ala	tgo Cys 350	: Val	g cgc Arg	1056
ato Ile	ccg	ato Ile	acc Thr	ggc Gly	ago Ser	aac Asr	ccg	J aac	g gcc s Ala	aag Lys	g egg	g cto	g gag ı Glı	g tto 1 Phe	c cga	1104

355 360 365 age eee gae teg teg gge aac eeg tat etg geg tte teg gee atg etg 1152 Ser Pro Asp Ser Ser Gly Asn Pro Tyr Leu Ala Phe Ser Ala Met Leu 375 370 380 atg gca ggc ctg gac ggt atc aag aac aag atc gag ccg cag gcg ccc 1200 Met Ala Gly Leu Asp Gly Ile Lys Asn Lys Ile Glu Pro Gln Ala Pro 390 395 385 1248 gte gae aag gat ete tae gag etg eeg eeg gaa gag gee geg agt ate Val Asp Lys Asp Leu Tyr Glu Leu Pro Pro Glu Glu Ala Ala Ser Ile 405 410 1296 ccq cag act ccg acc cag ctg tca gat gtg atc gac cgt ctc gag gcc Pro Gln Thr Pro Thr Gln Leu Ser Asp Val Ile Asp Arg Leu Glu Ala 420 425 gac cac gaa tac ctc acc gaa gga ggg gtg ttc aca aac gac ctg atc 1344 Asp His Glu Tyr Leu Thr Glu Gly Gly Val Phe Thr Asn Asp Leu Ile 440 435 1392 gag acg tgg atc agt ttc aag cgc gaa aac gag atc gag ccg gtc aac Glu Thr Trp Ile Ser Phe Lys Arg Glu Asn Glu Ile Glu Pro Val Asn 455 460 1437 ate egg eeg cat eec tae gaa tte geg etg tae tae gae gtt taa Ile Arg Pro His Pro Tyr Glu Phe Ala Leu Tyr Tyr Asp Val 470 475 <210> 8 0 <211> 478 <212> PRT <213> Mycobacterium tuberculosis <220> <221> <222> Sequence is identical to SwissProt entry SP:GLN1\_MYCTU <223> Sequence is identical to PIR entry PIR:H70775 Sequence is identical to PRF entry PRF:2323405A Met Thr Glu Lys Thr Pro Asp Asp Val Phe Lys Leu Ala Lys Asp Glu Lys Val Glu Tyr Val Asp Val Arg Phe Cys Asp Leu Pro Gly Ile Met Gln His Phe Thr Ile Pro Ala Ser Ala Phe Asp Lys Ser Val Phe Asp 35 40 45

Asp Gly Leu Ala Phe Asp Gly Ser Ser Ile Arg Gly Phe Gln Ser Ile

His	Glu	Ser	Asp	Met	Leu	Leu	Leu	Pro	qaA	Pro	Glu	Thr	Ala	Arg	Ile
65			_		70					75					80

- Asp Pro Phe Arg Ala Ala Lys Thr Leu Asn Ile Asn Phe Phe Val His
  85 90 95
- Asp Pro Phe Thr Leu Glu Pro Tyr Ser Arg Asp Pro Arg Asn Ile Ala 100 105 110
- Arg Lys Ala Glu Asn Tyr Leu Ile Ser Thr Gly Ile Ala Asp Thr Ala 115 120 125
- Tyr Phe Gly Ala Glu Ala Glu Phe Tyr Ile Phe Asp Ser Val Ser Phe 130 135 140
- Asp Ser Arg Ala Asn Gly Ser Phe Tyr Glu Val Asp Ala Ile Ser Gly 145 150 155 160
- Trp Trp Asn Thr Gly Ala Ala Thr Glu Ala Asp Gly Ser Pro Asn Arg 165 170 175
- Gly Tyr Lys Val Arg His Lys Gly Gly Tyr Phe Pro Val Ala Pro Asn 180 185 190
- Asp Gln Tyr Val Asp Leu Arg Asp Lys Met Leu Thr Asn Leu Ile Asn 195 200 205
- Ser Gly Phe Ile Leu Glu Lys Gly His His Glu Val Gly Ser Gly Gly 210 215 220
- Gln Ala Glu Ile Asn Tyr Gln Phe Asn Ser Leu Leu His Ala Ala Asp 225 230 235 240
- Asp Met Gln Leu Tyr Lys Tyr Ile Ile Lys Asn Thr Ala Trp Gln Asn 245 250 255
- Gly Lys Thr Val Thr Phe Met Pro Lys Pro Leu Phe Gly Asp Asn Gly 260 265 270
- Ser Gly Met His Cys His Gln Ser Leu Trp Lys Asp Gly Ala Pro Leu 275 280 285

Met Tyr Asp Glu Thr Gly Tyr Ala Gly Leu Ser Asp Thr Ala Arg His 290 295 300

Tyr Ile Gly Gly Leu Leu His His Ala Pro Ser Leu Leu Ala Phe Thr 305 310 315 320

Asn Pro Thr Val Asn Ser Tyr Lys Arg Leu Val Pro Gly Tyr Glu Ala 325 330 335

Pro Ile Asn Leu Val Tyr Ser Gln Arg Asn Arg Ser Ala Cys Val Arg 340 345 350

Ile Pro Ile Thr Gly Ser Asn Pro Lys Ala Lys Arg Leu Glu Phe Arg 355 360 365

Ser Pro Asp Ser Ser Gly Asn Pro Tyr Leu Ala Phe Ser Ala Met Leu 370 380

Met Ala Gly Leu Asp Gly Ile Lys Asn Lys Ile Glu Pro Gln Ala Pro 385 390 395 400

Val Asp Lys Asp Leu Tyr Glu Leu Pro Pro Glu Glu Ala Ala Ser Ile 405 410 415

Pro Gln Thr Pro Thr Gln Leu Ser Asp Val Ile Asp Arg Leu Glu Ala 420 425 430

Asp His Glu Tyr Leu Thr Glu Gly Gly Val Phe Thr Asn Asp Leu Ile 435 440 445

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Ile Arg Pro His Pro Tyr Glu Phe Ala Leu Tyr Tyr Asp Val 465 470 475

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<212> DNA

<213> Mycobacterium tuberculosis

<220>

<221> CDS

<222> (1)..(1341)
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of GenBank entry GB:MTCY427 [Z70692]
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teg gte gee ate gee cea gee gaa ete gag gge gee tte gag gaa gge Ser Val Ala Ile Ala Pro Ala Glu Leu Glu Gly Ala Phe Glu Glu Gly 35 40 45

atc ggc ttc gac gga tcc tcg atc gag ggc ttt gcg cgg gtc tcg gaa 192
Ile Gly Phe Asp Gly Ser Ser Ile Glu Gly Phe Ala Arg Val Ser Glu
50 60

tcc gat acg gtg gcg cac ccg gac ccg tcg acc ttc cag gtg ctg ccc
Ser Asp Thr Val Ala His Pro Asp Pro Ser Thr Phe Gln Val Leu Pro
65 70 75 80

tgg gcc acc agt tcc ggc cac cac cac tca gcg cgg atg ttt tgc gac

Trp Ala Thr Ser Ser Gly His His His Ser Ala Arg Met Phe Cys Asp

85 90 95

atc acc atg ccg gac ggc tcg ccg tcg tgg gcg gac ccg cgg cac gtg

Ile Thr Met Pro Asp Gly Ser Pro Ser Trp Ala Asp Pro Arg His Val

100 105 110

ttg cgg cgg cag ctg acg aag gcc ggc gaa ctc ggc ttc tcc tgc tac 384
Leu Arg Arg Gln Leu Thr Lys Ala Gly Glu Leu Gly Phe Ser Cys Tyr
115 120 125

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Val His Pro Glu Ile Glu Phe Phe Leu Leu Lys Pro Gly Pro Glu Asp
130 135 140

ggg tcg gtg ccc gtc ccg gtc gac aac gcc ggc tat ttc gac caa gcg
Gly Ser Val Pro Val Pro Val Asp Asn Ala Gly Tyr Phe Asp Gln Ala
145
150
150
160

gtg cac gac tcc gcc ttg aac ttt cgc cgc cac gcg atc gat gcc ctg
Val His Asp Ser Ala Leu Asn Phe Arg Arg His Ala Ile Asp Ala Leu
165 170 175

gaa ttc atg ggc atc tcg gtg gag ttc agc cat cac gaa ggc gca ccc 576 Glu Phe Met Gly Ile Ser Val Glu Phe Ser His His Glu Gly Ala Pro 180 185 190

ggc cag cag gag atc gac ctg cgg ttt gcc gac gct ctg tcg atg gct 624

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gag Glu 225	ggc Gly	gcc Ala	cgg Arg	gcg Ala	tcg Ser 230	ttc Phe	atg Met	ccc Pro	aag Lys	cca Pro 235	ttc Phe	ggc ggc	cag Gln	cac His	ccg Pro 240	720
ggc	tcg Ser	gcg Ala	atg Met	cac His 245	acc Thr	cac His	atg Met	agc Ser	ctg Leu 250	ttc Phe	gag Glu	ggt Gly	gat Asp	gtc Val 255	aac Asn	768
					gat Asp											816
tcg Ser	ttc Phe	atc Ile 275	gcc Ala	ej aaa	atc Ile	ctg Leu	gag Glu 280	cac His	gct Ala	tgc Cys	gag Glu	atc Ile 285	agc Ser	gcg Ala	gtc Val	864
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gcg Ala 305	ccc Pro	acg Thr	gcc Ala	gcg Ala	tcg Ser 310	tgg Trp	ggg ggg	gcc Ala	gcc Ala	aac Asn 315	cga Arg	tcc Ser	gcc Ala	cta Leu	gtg Val 320	960
cgg Arg	gtg Val	ccg Pro	atg Met	tac Tyr 325	acg Thr	ccg Pro	cac His	aag Lys	acc Thr 330	tcg Ser	tcg Ser	cgg Arg	cgg Arg	gtc Val 335	gaa Glu	1008
gta Val	cgc Arg	agc Ser	cct Pro 340	gat Asp	tcg Ser	gcg Ala	tgc Cys	aat Asn 345	Pro	tat Tyr	ctg Leu	aca Thr	ttc Phe 350	Ala	gtg Val	1056
ctg Leu	ctg Leu	gcc Ala 355	Ala	gga Gly	ttg Leu	Arg	ggt Gly 360	Val	gag Glu	aag Lys	ggt Gly	tac Tyr 365	Val	ctg Leu	ggc	1104
ccg Pro	cag Gln 370	Ala	gag Glu	gac Asp	aac Asn	gta Val 375	Trp	gac Asp	ctc Leu	aca Thr	Pro 380	Glu	gaa Glu	cgc Arg	cga Arg	1152
gcg Ala 385	Met	gly	tac Tyr	cga Arg	gaa Glu 390	Leu	ccg Pro	tco Ser	agt Ser	ttg Leu 395	Asp	agt Ser	gcg Ala	ctg Leu	cgc Arg 400	1200
gcc Ala	atg Met	gag Glu	gcc Ala	tcc Ser 405	Glu	ctc Leu	gto Val	gcg Ala	gag Glu 410	. Ala	ttg Lev	. <b>G</b> 17	gag Glu	cac His 415	gtt Val	1248
															cgc Arg	1296

Πij

420 425 430

agc cac gtc acg cca tac gag ctg cgc acc tac ctg tcg ctg tag Ser His Val Thr Pro Tyr Glu Leu Arg Thr Tyr Leu Ser Leu 435 440 445 1341

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<213> Mycobacterium tuberculosis

<220>

<221>

<222>

<223> Sequence is identical to SwissProt entry SP:GLN2\_MYCTU Sequence is identical to PIR entry PIR:B70776

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Ile Gly Phe Asp Gly Ser Ser Ile Glu Gly Phe Ala Arg Val Ser Glu 50 55 60

Ser Asp Thr Val Ala His Pro Asp Pro Ser Thr Phe Gln Val Leu Pro 65 70 75 80

Trp Ala Thr Ser Ser Gly His His Ser Ala Arg Met Phe Cys Asp 85 90 95

Ile Thr Met Pro Asp Gly Ser Pro Ser Trp Ala Asp Pro Arg His Val

Leu Arg Arg Gln Leu Thr Lys Ala Gly Glu Leu Gly Phe Ser Cys Tyr 115 120 125

Val His Pro Glu Ile Glu Phe Phe Leu Leu Lys Pro Gly Pro Glu Asp 130 135 140

Gly Ser Val Pro Val Pro Val Asp Asn Ala Gly Tyr Phe Asp Gln Ala

 Val His Asp Ser Ala Leu Asn Phe Arg Arg His Ala Ile Asp Ala Leu 165 170 175

Glu Phe Met Gly Ile Ser Val Glu Phe Ser His His Glu Gly Ala Pro 180 185 190

Gly Gln Gln Glu Ile Asp Leu Arg Phe Ala Asp Ala Leu Ser Met Ala 195 200 205

Asp Asn Val Met Thr Phe Arg Tyr Val Ile Lys Glu Val Ala Leu Glu 210 215 220

Glu Gly Ala Arg Ala Ser Phe Met Pro Lys Pro Phe Gly Gln His Pro 225 230 235 240

Gly Ser Ala Met His Thr His Met Ser Leu Phe Glu Gly Asp Val Asn 245 250 255

Ala Phe His Ser Ala Asp Asp Pro Leu Gln Leu Ser Glu Val Gly Lys 260 265 270

Ser Phe Ile Ala Gly Ile Leu Glu His Ala Cys Glu Ile Ser Ala Val 275 280 285

Thr Asn Gln Trp Val Asn Ser Tyr Lys Arg Leu Val Gln Gly Glu 290 295 300

Ala Pro Thr Ala Ala Ser Trp Gly Ala Ala Asn Arg Ser Ala Leu Val 305 310 315 320

Arg Val Pro Met Tyr Thr Pro His Lys Thr Ser Ser Arg Arg Val Glu 325 330 335

Val Arg Ser Pro Asp Ser Ala Cys Asn Pro Tyr Leu Thr Phe Ala Val 340 345 350

Leu Leu Ala Ala Gly Leu Arg Gly Val Glu Lys Gly Tyr Val Leu Gly 355 360 365

Pro Gln Ala Glu Asp Asn Val Trp Asp Leu Thr Pro Glu Glu Arg Arg 370 375 380

Ala M 385	iet (	3ly '	Tyr	Arg	Glu 390	Leu	Pro	Ser		Leu 395	Asp	Ser	Ala	Leu	Arg 400	
Ala M	let (	3lu :	Ala	Ser 405	Glu	Leu	Val	Ala	Glu 410	Ala	Leu	Gly	Glu	His 415	Val	
Phe 7	lsp :		Phe 420	Leu	Arg	Asn	Lys	Arg 425	Thr	Glu	Trp	Ala	Asn 430	Tyr	Arg	
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gag Glu	ggc Gly	gtc Val	gac Asp 20	acc	gto Val	atc Ile	ggc	acc Thr 25	gtc Val	gtg Val	aac Asr	e cco	gcc Ala 30	gga Gly	ctc Leu	96
acc Thr	cag Gln	gcc Ala 35	aag Lys	acc Thr	gtg Val	ccg Pro	ata Ile 40	. cgc : Arg	cgg Arg	acc Thr	aac Asr	aca Thi 45	tto Phe	gcc Ala	aat Asn	144
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ctc Leu	999	atc	gat	cto	~ +~	a acc	tte	cac	ato	ato	e aa	da da	a a	a tte	- ~~~	288

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atg Met	ctg Leu	tto Phe	s tcc Ser 260	: Gly	gly aaa	act Thr	gga Gly	gca Ala 265	. Ala	ggq Gly	ato Met	g acc	Ser 270	Ala	e GJA s aaa	816
gag Glu	gco Ala	gcg Ala 275	a Val	gca . Ala	gga Gly	gtg Val	Lev 280	Arg	gly gga	a cta / Le:	a ccc	g gad Asj 28!	, Ala	c caa a Gli	a ggc	864
ato Ile	ctg Leu 290	ι Суя	gga Gl	tcg Ser	ato : Ile	gto Val	. Sei	ggt Gly	cto Lei	g cga	a atg g Mei 300	t Ar	a cco	e gg	t aac y Asn	912
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gag gtg aag gtc gtc gac ccg tcg gcc aac ccg tat ctc gcg tcg gcg Glu Val Lys Val Val Asp Pro Ser Ala Asn Pro Tyr Leu Ala Ser Ala 340 345 350	1056
gcg atc ctc gga ctg gca ctc gac ggc atg aag acc aag gcg gtg ttg Ala Ile Leu Gly Leu Ala Leu Asp Gly Met Lys Thr Lys Ala Val Leu 355 360 365	1104
ccg tcg gaa acg acc gta gac ccg aca cag ctg tct gac gtg gat cgt Pro Ser Glu Thr Thr Val Asp Pro Thr Gln Leu Ser Asp Val Asp Arg 370 375 380	1152
gac cgt gcc ggc att ctg cga ctt gct gcc gat cag gcg gat gca att Asp Arg Ala Gly Ile Leu Arg Leu Ala Ala Asp Gln Ala Asp Ala Ile 385 390 395 400	1200
gct gta ctg gat agt tcg aaa ctg ctt cgg tgc atc ctt ggc gat ccc Ala Val Leu Asp Ser Ser Lys Leu Leu Arg Cys Ile Leu Gly Asp Pro 405 410 415	1248
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ggt gac ctc gat cct gcg cag ctg gcc gac aag ttc cgg atg gct tgg Gly Asp Leu Asp Pro Ala Gln Leu Ala Asp Lys Phe Arg Met Ala Trp 435 440 445	1344
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Pro Gly Leu Gly Ala Ser Pro Val Trp His Thr Phe Cys Ile Asp Gln 50 55 60

Cys Ser Ile Ala Phe Thr Ala Asp Ile Ser Val Val Gly Asp Gln Arg 65 70 75 80

Leu Arg Ile Asp Leu Ser Ala Leu Arg Ile Ile Gly Asp Gly Leu Ala 85 90 95

Trp Ala Pro Ala Gly Phe Phe Glu Gln Asp Gly Thr Pro Val Pro Ala 100 105 110

Cys Ser Arg Gly Thr Leu Ser Arg Ile Glu Ala Ala Leu Ala Asp Ala 115 120 125

Gly Ile Asp Ala Val Ile Gly His Glu Val Glu Phe Leu Leu Val Asp 130 135 140

Ala Asp Gly Gln Arg Leu Pro Ser Thr Leu Trp Ala Gln Tyr Gly Val 145 150 155 160

Ala Gly Val Leu Glu His Glu Ala Phe Val Arg Asp Val Asn Ala Ala 165 170 175

Ala Thr Ala Ala Gly Ile Ala Ile Glu Gln Phe His Pro Glu Tyr Gly
180 185 190

Ala Asn Gln Phe Glu Ile Ser Leu Ala Pro Gln Pro Pro Val Ala Ala 195 200 205

Ala Asp Gln Leu Val Leu Thr Arg Leu Ile Ile Gly Arg Thr Ala Arg 210 215 220

Arg His Gly Leu Arg Val Ser Leu Ser Pro Ala Pro Phe Ala Gly Ser 225 230 235 240

Ile Gly Ser Gly Ala His Gln His Phe Ser Leu Thr Met Ser Glu Gly 245 250 255

Met Leu Phe Ser Gly Gly Thr Gly Ala Ala Gly Met Thr Ser Ala Gly 260 265 270

Glu Ala Ala Val Ala Gly Val Leu Arg Gly Leu Pro Asp Ala Gln Gly 275

Ile Leu Cys Gly Ser Ile Val Ser Gly Leu Arg Met Arg Pro Gly Asn

Trp Ala Gly Ile Tyr Ala Cys Trp Gly Thr Glu Asn Arg Glu Ala Ala 310

Val Arg Phe Val Lys Gly Gly Ala Gly Ser Ala Tyr Gly Gly Asn Val 330 325

Glu Val Lys Val Val Asp Pro Ser Ala Asn Pro Tyr Leu Ala Ser Ala 350

Ala Ile Leu Gly Leu Ala Leu Asp Gly Met Lys Thr Lys Ala Val Leu 365 355 360

Pro Ser Glu Thr Thr Val Asp Pro Thr Gln Leu Ser Asp Val Asp Arg 370 375

Asp Arg Ala Gly Ile Leu Arg Leu Ala Ala Asp Gln Ala Asp Ala Ile 385 390

Ala Val Leu Asp Ser Ser Lys Leu Leu Arg Cys Ile Leu Gly Asp Pro 405

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cgc Arg	cgt Arg 130	Gln	ctc Leu	gat Asp	cgg Arg	ctc Leu 135	Ьys	gcg Ala	cgc Arg	gga Gly	ctg Leu 140	gtc Val	gcc Ala	gat Asp	gtg Val	4	132
gcc Ala 145	Thr	gag Glu	ctg Leu	gag Glu	ttc Phe 150	Ile	gtg Val	ttc Phe	gac Asp	cag Gln 155	Pro	tat Tyr	cgc Arg	cag Gln	gca Ala 160	4	180
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Ile	Arg	Leu 195	Gly	Met	Ala	Gly	Ala 200	Gly	Leu	Arg	Phe	Glu 205	Ala	Val	Lys	
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~ ~	_	_		_	gac Asp 230			_			_			_	_	720
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					cgg											1056
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420 425 430

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Asp Asp Ile Ala Thr Arg Gly Val Glu Cys Cys Ser Tyr Leu Leu Ala 50 55 60

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Asp Thr Gly Tyr Gly Asp Met Val Met Thr Pro Asp Leu Ser Thr Leu 85 90 95

Arg Leu Ile Pro Trp Leu Pro Gly Thr Ala Leu Val Ile Ala Asp Leu 100 105 110

Val Trp Ala Asp Gly Ser Glu Val Ala Val Ser Pro Arg Ser Ile Leu 115 120 125

Arg Arg Gln Leu Asp Arg Leu Lys Ala Arg Gly Leu Val Ala Asp Val 130 135 140

Ala Thr Glu Leu Glu Phe Ile Val Phe Asp Gln Pro Tyr Arg Gln Ala 145 150 155 160

Trp Ala Ser Gly Tyr Arg Gly Leu Thr Pro Ala Ser Asp Tyr Asn Ile 165 170 175

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Ala Leu Val Thr Cys Asp Asn His Ala Ile Tyr Lys Asn Gly Ala Lys 225 230 235 240

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Asp Glu Arg Glu Gly Asn Ser Cys His Ile His Val Ser Leu Arg Gly 260 265 270

Thr Asp Gly Ser Ala Val Phe Ala Asp Ser Asn Gly Pro His Gly Met 275 280 285

Ser Ser Met Phe Arg Ser Phe Val Ala Gly Gln Leu Ala Thr Leu Arg 290 295 300

Glu Phe Thr Leu Cys Tyr Ala Pro Thr Ile Asn Ser Tyr Lys Arg Phe 305 310 315 320

Ala Asp Ser Ser Phe Ala Pro Thr Ala Leu Ala Trp Gly Leu Asp Asn 325 330 335

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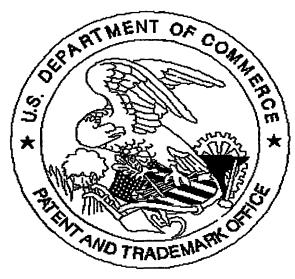
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Glu Arg Ile Arg Gly Phe Glu Arg Leu 450 455

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